Effects of Compost on Nematode *Pratylenchus* sp. (Tylenchida: Pratylenchidae) Population in Patchouli

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ABSTRACT

Patchouli is seen as one of the most important export commodities in Indonesia. The presence of parasitic nematode *Pratylenchus* sp. creates a barrier to plant cultivation. Increasing organic content in soil by applying compost in the plantation is one of approaches in controlling the nematode. This research was mainly aimed to study the effects of compost fertilization on the population growth of nematode *Pratylenchus* sp. and yields on patchouli. These research used a Completely Randomised Design (CRD) consisting of 12 treatments with three replicates. Selected plant was inoculated with nematodes with the following initial population: 100, 200, 300, 400, and 500 nematodes in soil with and without compost. The plant with no inoculation was used as control. The result of the research confirmed that the addition of compost was capable of suppressing the population of nematode *Pratylenchus* sp. as much as 58.93 - 67.31% and 40.74 - 59.19% in root and in soil of each treatment, respectively. Compost applied in each treatment positively affected the increasing amount of Patchouli oil yielded, and it could also reduce the loss as much as 18.34 - 43.36%. *Trichoderma* sp. was found to be dominant in compost in the treatment with its population of 2.1×10^4 propogul/g.

Keywords: Organic matters, patchouli, plant-parasitic nematode, Trichoderma sp.

INTRODUCTION

Patchouli oil serves as basic, secondary, and fixative ingredients commonly used in perfumes, cosmetics, foods and drinks [1]. Yellow or red disease caused by parasitic nematode is to blame for the falling figures for patchouli essential oil production. This type of nematode is wide in range from *Pratylenchus* spp. (*Pratylenchus coffeae* and *Pratylenchus brachyurus*) which could wound the plant, *Meloidogyne* spp. (*Meloidogyne incognita* and *Meloidogyne hapla*) causing pustules on the plant, to the borer nematode called *Radopholus similis* [2].

Endoparasite such as nematode *Pratylenchus* spp., which widely spreads in Indonesia, massively infects patchouli. Despite some attempts such as introduction of nematicide, technical culture, and resistant cultivar, the results gained were not as expected. Although the

use of nematicide could help increase the productivity of plant up to 25%, the intensity of the application could negatively affect the quality of patchouli oil, environment, balance of ecosystems, and human health [3]. Seeing this fact, it is perceived that more organic steps need to be taken into account, one of which is by using compost [1]. This research was conducted to investigate the effects of compost fertilization on the development of population of nematode Pratylenchus sp. and yield of patchouli oil.

MATERIALS AND METHODS

The research was conducted in research field and Nematology Laboratory, Department of Pest and Plant Diseases, Faculty of Agriculture, Brawijaya University from October 2014 to April 2015. Prior to the research, growing media, nematode multiplication for inocula-

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tion, and plant samples were prepared.

Preparing growing media

Soil used in this research was obtained from Jatikerto Village, Kromengan District, Malang Regency. Soil was sterilised with formalin (4%) in anaerobic condition for three days. The sterilised soil was then transferred into polybags, each of which was filled with 5 kg soil. When ready, these polybags served as growing media for plants during the experiment.

Preparation and multiplication of nematode

Before inoculum was multiplied, two patchouli plant samples in polybags were prepared. Nematode Pratylenchus sp. inoculum was isolated from soil and root of coffee tree infested with nematode Pratylenchus sp. in Kadudampit District, Sukabumi Regency, East Java. The nematode was then extracted by using Baermann funnel [4], where 5 funnels would be required for root and soil sample. The soil and root were cleaned from any debris before the root was sliced off as long as 0.5 - 1 cm and weighed to obtain 10 g for a sample and 100 g for soil sample. Separately, the funnels were prepared by placing tissue on top of wire screen into each of the funnel. Each prepared sample was then placed into each funnel and watered. After 24 hours, the extract was obtained and transferred into bottles. The suspension with nematodes was poured into petri dish. The nematodes were taken by using inoculating needle and transferred onto a piece of glass that had been given a drop of glycerin formal fixative solution. Next, the glass was covered for observation and morphological identification. When the identification result showed the presence of Pratylenchus sp., the nematode was to be multiplied by pouring the suspension around the patchouli.

Plant sample preparation for experiment

The Patchouli was obtained from the nursery located in Kesamben District, Blitar Regency. It was multiplied by applying shoot-cutting taken from young but woody branches of trees with the length of 10-20 cm, and it was then planted in prepared sand. When it was 1.5 month old and turned into a seed, it was ready to be transferred into the prepared polybags, where each was planted with one seed of patchouli, and 31 g compost was applied when planting.

Effects of compost on population of nematode Pratylenchus sp.

This research employed Completely Randomised

Design (CRD) which comprised 6 stages of initial population of nematodes: 0, 100, 200, 300, 400, and 500 nematodes and two growing media such as soil with and without compost. The initial numbers of population of nematodes and growing media were combined and 12 treatments were obtained. Each treatment was replicated three times which made up 36 treatments in total. The treatments tested are as follows:

| А | : 0 nematode + soil |
|---|----------------------------------|
| В | : 0 nematode + soil + compost |
| С | : 100 nematodes + soil |
| D | : 100 nematodes + soil + compost |
| Е | : 200 nematodes + soil |
| F | : 200 nematodes + soil + compost |
| G | : 300 nematodes + soil |
| Н | : 300 nematodes + soil + compost |
| Ι | : 400 nematodes + soil |
| J | : 400 nematodes + soil + compost |
| Κ | : 500 nematodes + soil |
| L | : 500 nematodes + soil + compost |

Pratylenchus sp. was prepared from extract obtained from the root of coffee tree infested with the nematodes, while the multiplication was obtained by using Baermann funnel method as explained above. The extract in the form of suspension was transferred into a bottle before it was poured into Petri dish. From the Petri dish, the nematodes were taken by using inoculating needle and counted with hand counter. The nematodes would need to be categorized by their number of population: 100, 200, 300, 400, and 500 before they were placed on petri dishes separated based on the population number. Moreover, the counted nematodes were transferred into bottles also based on their category of population. Suspension with nematodes was poured around the ten-day-after-planting patchouli with the distance of about 5 cm from the plant and 3 cm in depth. This experimental plant was taken care of by weeding, tilling, controlling pest, and regular watering.

Total oil

Number of patchouli oil is done by steam distillation method using patchouli oil distillation equipment in Toxicology Laboratory, Department of Plant Pest, Brawijaya University.

Yield loss

To determine the yield losses due to nematode attttack calculated using the formula Chiarappa (1971 in Rival, 2006):

$$Y = X_0 - X_{100,....,500}$$

(1)

 Y
 : yield loss

 X₀
 : result of the control plants

 X_{100,...,500}
 : result in plants inoculated at various treatment initial population of nematodes

To obtain the percentage reduction in yield, is then calculated by the formula Harappa (1971 in Rival, 2006):

$$Z = (X_0-X_(100,...,500))/X_0 \times 100 \%$$
(2)
$$Z \qquad : percentage of yield loss$$

$$X_0 \qquad : heavy on control plants$$

$$X_{100,...,500} \qquad : weight in plants inoculated at various treatment initial population of nematodes$$

Percentage of suppression

To determine the percentage of suppression is calculated by the number of nematodes in the media without the compost reduced by the number of nematodes in the media with compost divided by the number of nematodes in the media without compost multiplied by one hundred percent.

Statistic analysis

Observational data obtained were analyzed by analysis of variance (F test) at 5% level to determine the effffect of treatment. If the result is significantly different then continued with Duncan Multiple Range Test at the 5% significance level to determine differences between treatments.

RESULTS AND DISCUSSION

Number of nematodes Pratylenchus sp. in root and soil

Averagely, the number of *Pratylenchus* sp. in root given treatment was significantly higher than that of control. The average population of nematodes *Pratylenchus* sp. was found higher (69.33) in the treatment of 500 nematodes + soil (Table 1).

The addition of compost was able to suppress the population of *Pratylenchus* sp. with the percentage ranging from 58.93 to 67.31%. The compost application seemed to be able to increase the activity of antagonistic fungal agents capable of colonizing plant roots, creating barrier for nematodes to penetrating the root tissues. As a result, their access to wound the plants was reduced. According to Harni and Mustika (2000), suggested that organic matters applied to soil could

suppress the population of *Pratylenchus* sp. in both root of patchouli and soil [5]. This application was also believed to be able to reduce the attack of nematode. This was due to the fact that the decomposed organic matters were toxic to nematode in addition to their ability to increase the activity of antagonistic microorganisms in soil.

Based on the identification, *Trichoderma* sp. was found dominant in compost given to treatments, with the population 2.1×10^4 propogul/g. According to Olabiyi (2013), stated that the use of compost composed of dead leaves mixed with *Trichoderma harzianum* Persoon significantly reduced the population of nematode *Pratylenchus* sp., *Xiphinema* sp., and *Meloidogyne* sp. with its parasitic ability [6], *Trichoderma* sp. was capable of directly reducing the possibility of disease and indirectly competing for space and nutrients.

The content of Trichoderma sp. in compost is believed to be able to produce enzymes and some secondary metabolite compounds which could suppress the population growth of the nematode and increase plant resistance to nematode attack. Those enzymes are able to control nematode [7]. Trichoderma sp. produces enzyme such as protease and chitinase which can degrade and rapture the egg's wall of nematodes. Besides, this fungal agent could also produce other enzymes such as peroxide, polyphenol oxidase, phenylalanine ammonia lyase which could induce systematic resistance of a plant. Sharon et al. (2001), confirmed that all strains of Trichoderma showed potential to colonize egg and second stage juvenile of Meloidogyne javanica in sterile in vitro test and to penetrate egg mass [8]. In short, the role of protease from Trichoderma spp. performs similar function to nematicide.

The increase in population of *Pratylenchus* sp. In 10 g root of patchouli and 100 g soil was in line with the increase of stage in initial population of nematode (Fig. 1). The increasing number of the nematodes in 10 g root and 100 g soil without compost was relatively higher than that in 10 g root and 100 g soil with compost.

The number of *Pratylenchus* sp. in root of patchouli planted in the medium without compost showed relatively higher increase than that in root planted in the medium added with compost. The average number of nematodes in root planted in soil added with compost was lower than that in root planted in just soil. According to Yedidia *et al.* (2000), asserted that *Trichoderma* sp. was potential to directly enhance plant growth and suppress the growth of pathogens by

| Treatment | Number of Nematodes in 10 g 100t (x ± SD) | Suppression (%) | Number of Nematodes in 100 g soil (x ± SD) | Suppression (%) |
|--------------------------------|--|-----------------|---|-----------------|
| 0 Nematode + Soil | 0.00 ± 0.00^{d} | | $0.00 \pm 0.00^{\circ}$ | |
| 0 Nematode + Soil + Compost | 0.00 ± 0.00^{d} | | $0.00 \pm 0.00^{\circ}$ | |
| 100 Nematodes + Soil | 37.33 ± 8.05^{abcd} | | 46.33 ± 3.68^{abc} | |
| 100 Nematodes + Soil + Compost | 15.33 ± 1.25^{bcd} | 58.93 | 23.67 ± 2.05^{bc} | 48.92 |
| 200 Nematodes + Soil | 31.33 ± 9.28^{abcd} | | 54.00 ± 9.27^{ab} | |
| 200 Nematodes + Soil + Compost | 12.67 ± 3.86^{cd} | 59.57 | 32.00 ± 2.94^{bc} | 40.74 |
| 300 Nematodes + Soil | 49.33 ± 9.84^{abc} | | 67.67 ± 10.65^{ab} | |
| 300 Nematodes + Soil + Compost | 16.67 ± 2.05^{bcd} | 66.21 | 34.00 ± 5.72^{bc} | 49.75 |
| 400 Nematodes + Soil | 57.33 ± 21.45^{ab} | | 72.33 ± 4.99^{ab} | |
| 400 Nematodes + Soil + Compost | 19.67 ± 1.70^{bcd} | 65.70 | 32.00 ± 2.45^{bc} | 55.76 |
| 500 Nematodes + Soil | 69.33 ± 2.62^{a} | | 90.67 ± 5.31^{a} | |
| 500 Nematodes + Soil + Compost | 22.67 ± 3.40^{bcd} | 67.31 | 37.00 ± 1.63^{bc} | 59.19 |

Table 1. The average population of Nematodes Pratylenchus sp. in 10 g root and 100 g soil with initial population of nematodes

Table 2. Average amount of patchouli oil and yield loss in several treatments with initial population of nematodes

| Treatment | Amount of patchouli oil (g/25 g dry terna) (x ± SD) | Oil yield loss (g/25 g dry terna) (x ± SD) | Yield loss in percentage (%) |
|--------------------------------|--|---|------------------------------|
| 0 Nematode + Soil | 1.15 ± 0.12^{ab} | $0.00 \pm 0.00^{\circ}$ | 0.00 |
| 0 Nematode + Soil + Compost | 1.57 ± 0.05 ^a | 0.00 ± 0.00 ^c | 0.00 |
| 100 Nematodes + Soil | 0.95 ± 0.06 abc | 0.20 ± 0.17 bc | 15.85 |
| 100 Nematodes + Soil + Compost | 1.27 ± 0.05^{a} | 0.30 ± 0.02^{bc} | 18.93 |
| 200 Nematodes + Soil | 0.92 ± 0.03^{abc} | 0.22 ± 0.09^{bc} | 18.91 |
| 200 Nematodes + Soil + Compost | 1.14 ± 0.05^{ab} | 0.42 ± 0.07^{abc} | 26.95 |
| 300 Nematodes + Soil | 0.42 ± 0.11^{bcd} | 0.73 ± 0.23^{ab} | 62.24 |
| 300 Nematodes + Soil + Compost | $0.88 \pm 0.05^{\text{abcd}}$ | 0.69 ± 0.10^{abc} | 43.90 |
| 400 Nematodes + Soil | 0.28 ± 0.16^{cd} | 0.86 ± 0.08^{ab} | 76.34 |
| 400 Nematodes + Soil + Compost | 0.79 ± 0.03^{abcd} | 0.77 ± 0.04^{ab} | 49.34 |
| 500 Nematodes + Soil | 0.07 ± 0.04^{d} | 1.08 ± 0.08^{a} | 94.16 |
| 500 Nematodes + Soil + compost | 0.77 ± 0.11^{abcd} | 0.80 ± 0.16^{ab} | 50.80 |

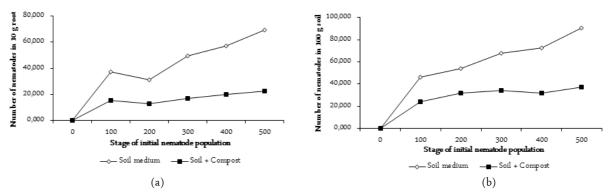


Figure 1. Correlation between stages of initial population of nematodes and average number of *Pratylenchus* sp. in 10g root of patchouli in two planting media (a) and in 100g soil in two planting media (b)

colonizing the area of rhizosphere and invading superficial layer of cortex of root, which reduced chambers for pathogens [9].

Similarly, the average number of *Pratylenchus* sp. In 100 g soil in several treatments was significantly higher than that of control. The highest average number of *Pratylenchus* sp. (90.67) was found in plant growing in soil + 500 nematodes.

Addition of compost in treatments could suppress the population growth of nematode Pratylenchus sp. In 100 g soil with suppression percentage ranging from 40.74 - 59.19%. The compost added in this experiment seemed able to increase organic content in soil and increase the population of antagonistic fungal agent such as Trichoderma sp. which could reduce the number of nematodes in soil. Dropkin (1996), stated that organic matters could be used to increase the population of antagonistic fungal agents, for compounds were released during the process of decomposition [10]. Decomposing remnants from plant tissues give off ammonia, salts, and simple organic acids such as acetic acid, propionate, and butyrate, which are toxic to nematodes. Organic matters added will be decomposed by microorganisms, forming and releasing toxic compounds which kill nematodes. According Dunn (1994), agreed that decomposition of organic matters served as nutrients to soil microbes such as fungi, Actynomycetes, and bacteria [11]. The increasing number of natural enemies for parasitic nematodes is able to control the population of nematodes naturally.

In addition, some secondary metabolites emitted by *Trichoderma* sp. were also toxic to nematodes' eggs, meaning that *Trichoderma* sp. also had potential to suppress the hatching. Stoppacher *et al.* (2010), reported that *Trichoderma* spp. was capable of producing volatile compound from its secondary metabolite [12]. According to Wheatley *et al.* (1997) and Yang *et al.* (2012), asserted that volatile compound determined the habitat of organisms. Volatile compound produced by *Trichoderma* sp. comprises 2-propanones, 2-methyl-1-butanols, heptane, octane and decane with their antimicrobial potential. Some other compounds produced also include 1β -vinylcyclopentane- 1α , 6-pentyl-2H-pyran-2-one, and 4-(2-hydroxyethyl) phenol, which are volatile and have potential to kill nematodes [13, 14].

Amount of oil and yield loss

The average amount of oil produced in several treatments was significantly lower than that in control group. The amount produced ranged from 0.07 - 1.57 g/25 g dry terna. Each plant infested with nematodes

of different initial population caused a decrease in the amount of oil yielded (Table 2).

The average amount of patchouli oil in plants with compost was relatively higher than that of plants without compost, so that the addition of compost positively influenced the increase in the amount of oil yielded.

The plant condition which was less resistant to the attack of the nematode seemed to affect the decrease of oil content yielded. It was obvious when the amount of oil kept decreasing as the initial of nematode population was increasing. Harni (1995), stated that the att-ttack of nematode could cause damage on plant and decrease up to 85% [15].

This decrease was believed to be caused by the damaged root, restricting it from absorbing sufficient nutrients and mineral which were required to help the plant grow and form oil content in patchouli. They were also needed in plant to form chlorophyll. When these elements are not presence, nitrogen (N), magnesium (Mg) and iron (Fe) will surely cause chlorosis.

The average oil yield loss in each treatment was higher than that of control, ranging from 0.20 - 1.08 g (15.85-94.16%). Harni (1995), reported that this loss due to nematode could reduce the yield up to 85% [15].

The table shows that the lowest yield loss (0.20 g) was obtained with the initial population of 100 nematodes + soil, while the highest (1.08 g) was with initial population of 500 nematodes + soil. The compost added in the treatment with initial population of 300-500 nematodes could reduce yield loss as much as 0.04-0.28 g (18.34 - 43.36%).

CONCLUSION

Compost could suppress the population of nematode *Pratylenchus* sp. as much as 58.93-67.31% and 40.74-59.19% in 10 g root of patchouli and in 100g soil in each treatment, respectively. The addition of compost in the treatments positively affected the increase in the amount of patchouli oil yielded, and it helped reduce the loss up to 18.34 - 43.36%.

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