

An Overview of Traditional Medicinal Plants Used in Treating Hepatocellular Carcinoma (HCC) with Emphasis on Mechanisms of Action

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ABSTRACT

Cancer is an incurable fatal disease wreaking havoc on the population of countries either developed or developing. Hepatocellular carcinoma (HCC) is the fifth most frequent type of cancer having a poor prognosis. Globally, HCC management and treatment intervention using medicinal plants from generation to generation is rapidly increasing and remains an area of promising research. This review provided insight into medicinal plants' ethnomedicinal use, photochemistry and mechanisms of action on HCC exhibiting anti-cancer potentials subjected to scientific investigations. Furthermore, clinical, toxicity studies and detailed mechanisms of action would provide an understanding of the development of lead compounds in drug discovery and development for HCC.

Keywords: Cancer, Hepatocellular carcinoma, Medicinal plants, Mechanisms

Introduction

Cancer remains one of the leading causes of death worldwide, and its prevalence is unabated with increasing early diagnosis, clinical intervention and increased public knowledge of common risk factors of this disease [1, 2] Cancer is a polygenic and multifactorial disease where multiple gene mutations cause normal cells to become abnormal or uncontrollable in growth patterns, ultimately leading to the evasion of cell cycle regulation and programmed cell death [3].

The cancer disease process happens during development when the human cells grow old, get damaged, and die; then new cells capture their position. The breakdown of the orderly process results in abnormal or damaged cell growth and multiplication without control. These cells will

possibly form tumours, which are lumps of tissue and might be cancerous or not (benign). Tumours that are cancerous spread to or invade close by tissues and can travel in the body to distant places to form fresh tumours, a process known as metastasis [4]. Cancer is also known as a severe health predicament in all populations irrespective of social or wealth class. The global reaction to cancer has been irregular and discriminatory.

The trouble of neoplastic or cancer diseases is a considerable global health confront accounting for considerably high cases of deaths. In 2022, about 2 million new incidences of liver cancer were diagnosed in the US and an estimated 609,360 death cancer projections with the occurrence of 1,670 deaths/day [5]. These projections

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were based on the reported incidence of cancer and its mortality in 2018 and 2019, respectively, and do not account for the unknown effects of COVID-19 (coronavirus disease 2019) on cancer diagnoses and deaths [4]. The incidence of cancer in Uganda was about 32,617 cancers as reported in 2018 with 21,829 deaths [6].

Liver cancer is reported to be the second most common cause of death related to cancer globally after lung cancer [7]. In addition, hepatocellular carcinoma HCC is one of the most frequent cancers that affect the liver, and the incidence rate is 500,00 to 1 million new cases yearly [2]. In the United State of America, liver cancer is the second most frequent cause of death after cardiac disease [4]. Liver surgical resection and transplantation are the most common and proficient therapeutic strategies for treating HCC at an early stage. The current synthetic drugs are still not potent or effective in treating advanced HCC [8]. Hence, drug discovery and development must be aimed toward products having high efficiency and low toxicity for HCC.

Generally, cancer treatment and management strategies include surgery, radiotherapy, chemotherapy, and hormonotherapy. The relapse of cancer and side effects of conventional treatment procedures leading to reduced quality of life patients (QOL) [9] have been significant bane attributable to the strategies above in cancer treatment. However, there is an increase in demand for developing novel approaches and complementary therapies to cancer treatment. One critical complementary approach is using plant-derived products that have been in use thousands of years ago with characteristics of very low side effects in cancer treatment [10,11]. Plant-derived constituents, among other pharmacological importance, directly inhibit the proliferation of cancer cells and display broad activity that targets many pathways [12]. This review is thus aimed at providing comprehensive information on the need for different medicinal plants for treating HCC with some mechanism of action involved.

Hepatocellular Carcinoma (HCC)

Liver cancer is the foremost cause of death, which is cancer-related globally. Cancer-related death is worrisome, and data indicated a 4.7 % death rate for all cancer types. Liver cancer is positioned sixth in terms of incidence and ranks third position in death rate globally [13], as attested by

the International Agency for Research on Cancer. An exceedingly aggressive and most common class of liver cancer, hepatocellular carcinoma (HCC), occurs in the fifth position. The incidence of HCC is topographically dependent, with most cases taking place in developing countries. Eastern Asia and Sub-Saharan Africa are where the highest incidence of HCC Occurs [14]. In developed countries like the United States, concern about the growing incidence of HCC is reported in the latest analysis from the Surveillance Epidemiology End Results (SEER). The projection revealed that the incidence might climax by 2030, especially among Hispanics, Blacks, Whites and Asian Americans, respectively. In many cases, liver cancer is caused by long-term damage and scars from the liver cirrhosis emanating from viral infections (Virus B or C) or non-viral causes such as autoimmune diseases, non-alcoholic Fatty Liver Disease (NAFLD), inflammation of the liver (chronic), diabetes, obesity, smoking, alcohol consumption, exposure to aflatoxin and iron overload in the body (hemochromatosis). The report confirmed that some of the ways of preventing HCC include vaccination and antiviral treatments. Hepatitis B vaccination remains the best intervention to reduce the incidence of HCC in places where hepatitis B virus (HBV) is highly prevalent, as demonstrated in Taiwan [15].

Risk factors/causative agents of HCC

HCC risk factors or causative agents are multifactorial, including gender, age, ethnicity, and geographical regions [16]. Chronic HBV infection and fungal toxin aflatoxins B1(AFB1) from contaminated food, are the most common factors [17]. HBV infection (Hepatitis B Virus) is regarded as a significant threat factor. Others, for example, are long-standing alcohol abuse, iron overload (either inherited or common in Sub Sahara African), cigarette smoking, membranous obstruction of the inferior vena cava (MOIVC), cirrhosis, alcoholic fatty liver disease (AFLD), obesity, diabetes, non-alcoholic fatty liver disease (NAFLD), some environmental chemicals that are carcinogenic leads to the development of HCC.

Virus and HCC

The burden of the virus as causative to the pathogenesis of HCC cannot be overemphasised. HBV is a partially double-stranded DNA virus belonging to the genus of Avihepadnavirus of the

Hepadnaviriae family. Out of the 240 million HBV infections in the world, HBV accounts for 75-80% of HCC [18]. HBV genetic material gets fused into the human genome, which induces inflammation, p53 inactivation, and oxidative stress, causing hepatocarcinogenesis. The proliferation and loss of growth control, sustained cycles of necrosis and regeneration, and activation of various oncogenic pathways such as PI3K/Akt/STAT3 pathway and Wnt/ β -catenin all lead to genomic instability associated with HCC [16].

Carcinogens and HCC

Carcinogens are cancer-causing agents involved in the aetiology of HCC. Aflatoxin, tobacco smoking (4-aminobiphenyl and polycyclic aromatic hydrocarbons), vinyl chloride, arsenic, cadmium, lead, and nickel are carcinogens implicated when an individual is exposed to them, and they contribute to the pathogenesis of HCC [8]. For example, aflatoxin induces mutation in the P53 tumour suppressor gene and causes abnormal growth of the liver, resulting in HCC development [19]. Carcinogens could act independently or synergistically with the genetic material of viruses to cause DNA damage, thus inducing cirrhosis while contributing to HCC.

Alcoholic Fatty Liver Disease (AFLD)-Associated HCC

AFLD is a major cause of liver disease burden and is responsible for 30% of HCC-related deaths in the world [20]. The pathogenesis of AFLD resulting from the excessive intake of alcohol starts with hepatic injury from the fat build, progression to inflammation and scarring leading to HCC [16]. In the United States, the baseline of more than 14 drinks/week and seven drinks/week for men and women, respectively is responsible for AFLD [21]. Variations depend on factors such as gender, ethnicity, and genetics in determining the threshold level of alcohol that induces hepatotoxic effects.

Iron overload and associated HCC

Iron overload, known as iron accumulation in the liver, has emerged to be an essential risk factor for HCC pathogenesis. An iron level in the body exceeding 5 g is known as iron overload. Large amounts are released into the liver's cytoplasm when the safe level sequestration of metal is exceeded, resulting in the denaturing of protein and

hepatic damage, bringing about HCC development [22].

Pathogenesis of HCC

The carcinogenesis mechanisms or pathogenesis of the diverse risk factors are complicated to be separated from the procedures leading to cirrhosis. Mechanistically, signalling pathways of related cellular oncogenes activation, unstable genome and tumour suppressor genes deactivation vis a vis DNA mismatch repair defects and impaired chromosomal segregation, overexpression of growth and angiogenic factors, and telomerase activation all contribute to HCC development [23, 24]. The mechanisms of hepatocarcinogenesis for the various risk factors are shown below. Common risk factors are indicated using the same colour. In addition to these mechanisms, hepatitis B virus (HBV) and aflatoxin B1 affect the genome — HBV can integrate into the host genome HCV, hepatitis C virus, and aflatoxin B1 is a mutagen. On the other hand, overexpression, or upregulation of rate-limiting enzymes (such as phosphofructokinase, hexokinase, lactate dehydrogenase and pyruvate kinase) in glycolytic pathway and increase in glucose uptake by overexpression of GLUTs enzymes have been attributed to HCC (Figure 1).

Detection and diagnosis of HCC

HCC is rarely detected early, and its fatality is much within a few months of its diagnosis. Imaging through multiphase computed tomography (CT) and/or multiphase magnetic image (MRI) scans forms the basis of HCC diagnosis. HCC are non-symptomatic until the advanced stage; hence, clinical features are seldom helpful in its diagnosis. Hence, combining abdominal ultrasound and diagnostic biomarkers helps increase the specificity and sensitivity of screening tests for HCC.

Treatment of HCC

Treatment options in HCC depend on the disease progression and the patient's state. Surgical resection, local ablation with radiofrequency, transcatheter arterial chemoembolization (TACE), radioembolization and targeted systemic therapeutic like the use of sorafenib are some of the available interventions [25]. The primary goal of treatment is probably finding the cure or prolonging life considerably. Another important goal is improving the patient's quality of life. This is achievable through psychosocial support and

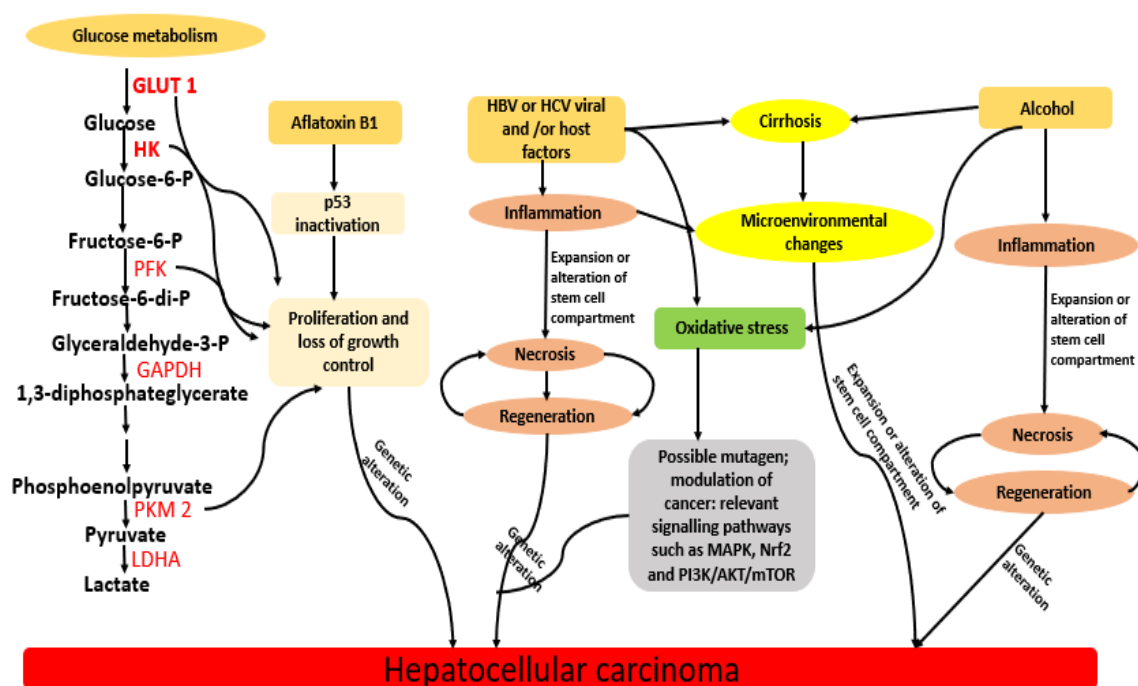


Figure 1. Different risk factors of HCC mechanisms or pathogenesis

supportive/palliative care. Multi-kinase inhibitors, for example, lenvatinib and sorafenib, are among the systemic drugs available for the treatment, and they are considered in patients with unresectable HCC cases.

Treatment through medicinal plant

Higher plants contribute more than 25% as natural products or derivatives out of the 50% of drugs already in use clinically [26]. Thus, plants remain the primary foundation of nearly all medicine in the world, which are important remedies for man. The knowledge of medicinal plants for their curative effects was accumulated over time from evidence-based observation [26]. In Africa, up to 80% of the population uses traditional medicines for Primary Health Care (PHC) (WHO, 2002). Adaptations of herbal medicines in developed countries are termed “Complementary” or “Alternative” Medicine [27,28]. In addition, to ascertain the importance of medicinal plants, 80% of the World’s population has benefited in healthcare needs from phytomedicinal sources, according to the estimation of the World Health Organization WHO [27], and even in the USA, about 20% of people have taken some herbal supplementation because approximately 25% of common medications contain herbs nowadays.

Promoting the use of herbal products in research and clinical settings requires knowledge about the pharmacokinetics and tissue distribution behaviour that is dependent on the physiological status of the body such as gender, age and disease state. In addition, oral bioavailability, tissue distribution, halftime ($t_{1/2}$), maximum plasma concentration (C_{max}), and time to reach C_{max} (T_{max}), are factors of pharmacokinetic parameters that cause variations in their therapeutic effects [29]. Herbal medicines include vegetable medicines, botanical medicines, or phytomedicines as described by WHO (World Health Organization)—some of the anticancer drugs available for treatment emanate from medicinal plants. Examples include vincristine and Vinblastine which are used as anticancer drugs that were discovered in vinca plants (*Catharanthus roseus*).

Research has shown that several medicinal plants have anticancer properties against a comprehensive variety of cancers, in particular HCC, with these anticancer properties accredited to the active chemical constituents available in these plants, as revealed in Table 1. Medicinal plants play a major role as therapeutic agents in traditional medicine contributing a major role in the treatment of hepatocellular carcinoma as mentioned in different literature reviews with their

Table 1. Traditional medicinal plants used in treating hepatocellular carcinoma (HCC) with their mechanism of action

PLANT	PLANT PART	EXTRACTION SOLVENT	PHYTOCHEMICALS	MECHANISM IN LIVER CANCERS	REFERENCES
<i>NephthiumLappaccum (Rambutan)</i>	Fruit	Aqueous Chloroform Ethyl acetate Hexane Methanol	Terpenoids, Alkaloids, Phenols, Anthocyanin, Carbohydrate, Triterpenoids, Cardiac glycosides, Coumarins, Flavonoids, Protein, Steroids, Saponins, Tannins,	cell proliferation controlling and caused shrinkage of HepG-2 cells spherical shape from polygonal	[37]
<i>Moringa Oleifera L.</i>	Leaf	Aqueous; Dichloromethane; Ethanol	saponins, alkaloid, cyanogenic glycoside and flavonoid	Decreasing cell proliferation and exhibiting apoptosis cell death in liver HepG-2	[38,6]
<i>Carica Papaya L.</i>	Leaf	Aqueous; n-hexane, Methanol	Alkaloids, Flavonoids, Saponins, Steroids, Tannins, Phlobatamine	Activate apoptosis by activating caspase 3/7 on Jurket cells	[39,6]
<i>Allium Sativum L.</i>	Bulb	Ethanol	Alkaloid, saponins, flavonoids, glycoside, anthraquinones, tamin and terpenoids	Increasing apoptosis by activating tumor suppressor (TP53) gene	[30,31,40,41,6]
<i>Abelmoschus Esculentus (L) Moench</i>	Fruit, Seed	Methanol	carbohydrates, sterols, flavonoids, proteins, tannins, terpenoids, phenols, and alkaloids	Proliferation and migration inhibition probably due to vascular endothelial growth factor (VEGF) production inhibition, leading to both apoptosis and death	[42,43,6]
<i>DioscoreaMembranacea</i>		Ethanol	saponins, phenanthrenes, anthocyanins, steroidal, saponins, diosgenin, and dioscin	Its decrease glypican3 and reticulon expression	[44,45]
<i>DracocephalumRiotschijBoiss (Labeatae)</i>	Aerial part	Ethanol, Methanol	Flavonoid, Luteolin	Induction of apoptosis via the opening of mitochondrial transmembrane pore	[46,47,31]
<i>EucomisAutumnalis (ML) Chitt</i>	Root	Methanol	Flavonoid, Phenolic acid, Eucomic acid	-	[48,36]
<i>Benja-ummarit</i>		Ethanol	Piperidine, Piperine, Piperetine, Dehydrodipiperidine, Piperidine	Inhibit cell production via vascular endothelial growth factor (VEGF) production inhibition	[49]
<i>Alisma Orientale</i>	Tuber	Methanol	Terpenoids, including sesquiterpenoids, diterpenoids, and triterpenoids	Increase apoptosis via down-regulating the Bax, p-JNK family protein and activation of Bcl-2 family proteins	[50,51]
<i>AristolochiaCucurbitifolia/macroura Go'mez</i>	Root, Stem	Methanol	Isoquinoline alkaloid and derivatives, phenanthrene derivatives, benzozate derivative	-	[51,53]
<i>Polygonum sp.</i>	Root		Resveratrol, flavonoids, quinones, phenylpropanoids, and terpenoids,	Resulting in apoptosis induction through the inhibition of integrins, cadherins and modulation of actin cytoskeleton organization	[51,54]
<i>Curtisia dentate (Burm F) C. A. Sn</i>	Leaves	Acetone	Lupeol, betulinic acid, ursolic acid and β-sitosterol	Involvement in the induction of apoptosis in HepG2 cell line	[55,36]

<i>Psidium Guajara</i>	Leaves	Aqueous, Methanol	Glycosides, Phenols, Saponins, and Tannins, Flavonoids, and Terpenoids	Proliferation of cells are inhibited in HepG2 cell line	[56,57,31,58]
Pacific Yew (<i>Taxus brevifolia</i>)	stem, leaf, and bark		bioflavonoids, lignans, phytoosterols, phytoecdysteroids.	–	[59]
<i>Marsdeniataenacissima</i>	Leaves, Root	Methanol	alkaloids, flavonoids, phenols, and saponins	Induces apoptosis by the up-regulation of caspase-9, Bax, and caspase-3 and inhibit anti-apoptotic which resulted to Bcl-2 and Bcl-XL expression down regulation. Is inhibit angiogenesis, promote apoptosis, improve immune function by targeting JAK-1, HIF1 α and P53, respectively.	[60,61,62]
<i>Coriolus versicolor</i> (Trametesversicolor) mushrooms		Aqueous, Methanol, Ethanol	Phenols, isoflavone molecule and flavonoids	Its suppresses proliferation of cells by suppressing the expression of Th1 cytokines IL-1 β , IL-2, TNF α , and IFN-1	[63,64,65]
<i>Basidiomycotina</i>		Water, hexane, butanol, methanol and ethyl acetate	phenolic and fatty acids	Increasing expression of proteins that is responsible for apoptosis and cell cycle arrest	[66,67]
<i>Viscum album L.</i> (mistletoe)			Flavonoid, Phenolic acid, Terpenoids, Lectin	It inhibits HCC cells proliferation through downregulation of c-Myc protein expression	[68,69]
<i>Panax ginseng</i>	Root, Leaves, Stem	Ethanol, Methanol	Ginsenosides, Phenolic, flavonoid, vitamin, Protopanaxadiol (PPD) Ginsenoside Rh2	Induction of anticancer activity via the instruction of apoptosis-related genes and the decrease the expression of cell cycle regulatory genes.	[70,71]
<i>Trametesrobiniophila</i>Murr mushrooms (Huaier granule)			Polysaccharides, steroids and alkaloids	It inhibits tumor cell proliferation, prevented metastasis, induced tumor cell death and interfered with angiogenesis via various signaling pathways.	[72,73,74]
<i>Epimediherba</i>			Flavonoids	It possesses anticancer activities via cell cycle regulation, apoptosis, angiogenesis, and metastasis, and a variety of signaling pathways including JAK2-STAT3, MAPK-ERK, and PI3k-Akt-mTOR.	[75,76]
<i>Camptotheca acuminata</i>	Leaves, Root, Fruit		fatty acids, alkaloids, flavonoids, polyphenols, tannins, terpenes, sterols and ellagic acids	Its antiproliferative activity is via induction of apoptotic proteins including Bcl-2 family protein	[77,78,79]
Poppy oil (<i>Papaver somniferum L</i>)	Seed		alkaloids, phenolic compounds, and essential oil	–	[80]
<i>Amaranthus spinosus</i>	Leaves, Stem	Ethanol, Methanol	Etlains, hydroxycinnamates, saponins, steroids and flavonoids	Inhibition of cell proliferation, induces apoptosis through down regulation of the expression of Bcl-2 with surviving while Bax, caspase-9, caspase-3, Apaf-1 and PARP were up regulated.	[81,82,79]

<i>Ziziphus jujube</i>	Ethanol	Ursolic acid, Oleanolic acid, Flavonols, saponins	Induce apoptotic effect and differential cell cycle apprehension in HepG2 cell	[83,84]
<i>Panax Ginseng</i>			Induces anti-proliferative effect, cell death and autophagy.	[85]
<i>Ardisia pusilla A. DC</i>	Methanol	Ardipulosilolide I, Triterpenoid, saponins, Phenylpropanoid, glycoside,	Inhibition the metastasis and invasion of HCC cell <i>in vivo</i> and <i>in vitro</i> via reduced metalloproteinase (MMP)-9 and MMP 2 proteins	[86,87,79,88]
<i>Digitalis ferruginea</i>	ethanol, acetone, water	LanatosideC, phenolic, flavonoid, tannin c	Inhibition of cell proliferation and induction of apoptosis and cell cycle arrest of G2/M phase by blocking MAPK/Wnt/PAM signaling pathways.	[89,90]
<i>Diospyros kaki</i>	Leaves	Non-specified terpenoids and flavonoids	induces cancer cell death and inhibits cell proliferation via PDGFR-Rac-JNK signaling axis is triggered by EEDK.	[91,92]
<i>Sanguisorba officinalis</i>	Methanol	phenolic acids, flavonoids, Ziyuglycoside II (3β-3-α-1- arabinopyranosyloxy)19-hydroxyurs-12- en-28-oicacid)	Inhibition of cell proliferation, apoptosis inducing and down-regulation of cell movement and invasion in various HCC cells via the EGFR/MAPK and EGFR/PI3K/AKT/NFκB signaling pathways	[93,94]
<i>Bruceajavanica</i>	Seed, Fruit	Brucejavanica oil, sesquiterpenes, olein, pregnane glucosides, oleic acid, anthraquinones, linoleic acid, and tetracyclic triterpene quassinoids	inhibit tumor cell from growing, affect the autophagy, apoptosis of tumor cell, invasion, and migration processes and activate immunity via NF-κB, p38-mitogen-activated protein kinase (MAPK), phosphoinositide-3-kinase (PI3K) protein kinase B (AKT), Wnt/β-catenin, and signal transducer and activator of transcription 3 (STAT3) pathways.	[31,95,96]
<i>Artemisia annua</i>		Artemisinin, coumarins, flavonoids and phenolic acid derivative	Promote anti- proliferative effect and induces apoptosis via reduction in activities of Akt/mTOR/anti-apoptotic proteins (like Bcl-XL and Bcl-2), that lead to increasing the activation of pro-apoptotic proteins (such as Bax and Bak) and tumor suppressor p53 via the Akt/mTOR signaling pathway.	[97,98]
<i>Citrus bergamia</i>	Fruit	Erriocitrin, Neoterocitrin, Naringin, Neohesperidin	Induces cell cycle arrest in G2 phase, activates the apoptotic machinery through both extrinsic and intrinsic pathways	[99,100]
<i>Scutellaria baicalensis</i>		Oroxylin A, baicalin, baicalein, wogonoside, and wogonin	Induction of G2/M cell cycle, activate and arrest caspase system on liver cell by ERK pathway upregulation	[101,102]

<i>Fagopyrum tataricum</i>	Tatariside F, flavonoids and phenolics	Induces antitumor effects in vivo and in vitro by up-regulating the protein expressions of p53 and Bax, and down-regulating the Bcl-2.	[103,79,104]
<i>Pulicariajaubertii</i>	Whole plant, aerial part	Pulicariajaubertii 1, hydroquinone, triterpenes	[105,106]
<i>Jujube (Ziziphus Jujuba)</i>	Leaves	terpenes, tannins, alkaloids, phenols, steroids, and cardiac glycosides	[79,107,108]
<i>Camellia sinensis</i>	Leaves	Epigallocatechin-3-gallate, alkaloids, flavonoids, steroids, terpenoids, carotenoids, benzoic acid, ascorbic acid, tocopherols, folic acid, and tannins consisting of catechin (flavonol) and gallic acids	[31,79,109]
<i>Garcinia mangostana</i>	Fruits	xanthenes and its derivatives (Mangostanaxanthone V, Mangostanaxanthone VI, Alpha-mangostin) and Flavonoid	[110,111,112]
<i>Rhizomacoptidis</i>		Berberine, alkaloids, volatile oils, organic acids, lignans, and flavones	[113,114,115]
<i>Nelumbo nucifera</i>	Seed	Neferine, alkaloids, flavonoids, sesquiterpenoids, essential oils	[116,117,118]
<i>Garcinia</i>	fruits, leaves, and seeds	Gambogic acid, anthones, flavonoids, phenolic acids, organic acids, and terpenoids	[119,120]
<i>Poncirus trifoliata</i>	Fruits	Poncirus fructus, Syringin, Citric acid, Poncirin	[121, 179]
<i>Alpinia galangal</i>	Rhizome, Leaves	10-Acetoxychavicol acetate, Phenolic, Flavonoid	[122,123]
White Birch Trees (<i>Betula papyrifera</i>)	Bark	Betulnic acid, Alkaloids, Phenols, Terpenoids	[79,124]
<i>Huanglian decoction</i>		CoptidisRhizoma, ZingiberisRhizoma, Folium ArtemisiaeArgyi, Mume Fructus, berberine, palmatine, baicalin and geniposide	[76,125]

<i>Allophylus cobbe</i>	Leaves, bark, Root, Stem	methanol, hexane, ethyl acetate, dichloromethane	Tannins, Phlobatannins, Steroids, Terpenoids, Cardiac glycosides, Flavonoids	[35,126,127]
<i>Madhuca longiflora</i>	Leaves, bark, Root, Stem	methanol, hexane, ethyl acetate, dichloromethane	sapogenins, triterpenoids, saponins, flavonoids and glycoside	[35,128]
<i>Adenanthera bicolor</i>	Leaves, bark	hexane, dichloromethane, ethyl acetate, and methanol	phenolic and total flavonoid	[35]
<i>Cyclea peltate</i>	Leaves, bark	hexane, dichloromethane, ethyl acetate, and methanol	tannin, proteins, resins, carbohydrates, alkaloids and terpenoids	[35,129,130]
<i>Munronia pinnata</i>	Leaves, bark	methanol, hexane, ethyl acetate, dichloromethane	Saponin, Alkaloids, Tannins, Flavonoids, and Steroid Glycosides	[35,131]
<i>Schumacheriacastaneifolia</i>	Leaves, bark	methanol, hexane, ethyl acetate, dichloromethane	sulphated flavonoids	[35,132]
<i>Carallia brachiata</i>	Leaves, bark	hexane, dichloromethane, ethyl acetate, and methanol	Alkaloids, Glycosides, Phenolic compounds and tannins, Flavonoids, Carbohydrates, Proteins, Fats and oils, Saponins	[35,133]
<i>Asparagus Racemosus</i>	Root	Aqueous	e steroidal saponins, essential oils, various amino acids, flavonoids, resin, steroidal tannin glycosides (asparaguses), and bitter glycosides	[134]
<i>Solanum nigrum</i>		Aqueous	alkaloids and steroid saponins glycoprotein, solasonine, steroidal glycosides, alpha, and beta solanigrinechez, alkaloidssolanine, margarine,	[134,135]
<i>Rubus Aleaifoliosus Poir</i>	Root	Butanol and ethyl acetate	Total alkaloids	[134, 180]
<i>Perilla frutescens</i> (commonly known as Korean perilla or Beefsteak vine)	Leaf		Phenolic acids, policosanol, flavonoids, triterpenes, phytosterols, carotenoids, fatty acids, tocopherols, and essential oils	[134,136]
<i>Ajwa Dates (Phoenix dactylifera L.)</i>		Methanol	flavonoids glycosides, polyphenol, and phytosterols	[134,137,138]
<i>Tumeric (Curcuma longa)</i>	Rhizome	Ethanollic	Curcumin, cyclocurcumin	[30,32,41,139]

Ginger essential oil	camphene, zingiberene, ocimene, z-citral, e-citral.		Induces relevant antiproliferative and antitumor activities by important mediators and pathways of cell signaling, including Bax/Bcl2, p38/MAPK, Nrf2, p65/NF- κ B, TNF- α , ERK1/2, SAPK/JNK, ROS/NF- κ B/COX-2, caspases-3, -9, and p53	[30,41,140]
<i>Saffron (Crocus sativus).</i>	carotene, crocin, anthocyanin, and lycopene.	Flower	Inhibits the cell proliferation via inducing apoptosis, modulation of oxidative damage and suppression of inflammatory response	[30,41,141]
<i>Artemisia vulgaris (Mugwort)</i>	flavonoids, coumarins, volatile oils, sesquiterpene, lactones, inulin, and traces of alkaloids	Aerial part		[30,142,143]
<i>Amorphophallus campanulatus (Elephant foot yam)</i>	Phenolic, tannins, sugar, starch, and total protein	Tuber	Induces cytotoxic and apoptotic activities	[30,144,145,146]
<i>Broussonetia luzonica</i>	carbohydrates, reducing sugars, flavonoids, tannins, alkaloids, and sterols	Leaves		[30,147]
<i>Graptopetalumparaguayense</i>	gallic acid, flavone, genistin, daidzin, and quercetin	Stem	Induced apoptosis in HCC cells by promoting the loss of mitochondrial membrane potential and the production of reactive oxygen species.	[30,148,149]
<i>Nigella sativa (Black seed or black cumin)</i>	monoterpene, di-terpene, sesquiterpenes, monoterpene alcohol, and ketone	Seed	Induces apoptosis and controls Akt pathway.	[30,150,151]
<i>Petasites japonicas</i>	Flavonoids, sesquiterpenes, triterpenes, phenol	Root	Anti-proliferative activity through inhibiting the Akt/mTOR and Wnt signaling pathways	[30,152,153]
<i>Asafoetida (Ferulaasafetida)</i>	farnesiferols A, B and C, asaresinotannols, coumarin derivatives (e.g. umbelliferone), ferulic acid, coumarin-sesquiterpene complexes (e.g. asacoumarin A and asacoumarin B)		induction of apoptosis and altered TGF- β and NF- κ B signaling with increase in TNF- α and caspase-3 expression	[32,154]
<i>Cinnamomum (Cinnamomum sp.)</i>	2-methoxycinnamaldehyde and cuminaldehyde	dry bark and twigs		[32]
<i>Dandelion (Taraxacum officinale)</i>	α -amyrin, β -amyrin, lupeol and taraxasterol	Root	Anti-proliferative by enhanced phosphorylation level of AMPK	[32,155]

Pomegranate (<i>Punica granatum</i> L.)	Leaf, fruit, peel, seed	Petroleum ether, dichloromethane, ethyl acetate, methanol and water	anthocyanins, 3-glucosides, cyanidin, pelargonidin, ellagitannins, delphinidin 3,5-diglucosides and other phenolic compounds, flavonoids, proanthocyanidin compounds, gallic acid, minerals and complex polysaccharides, ellagic acid and ethyl brevifolin-carboxylate, triterpene acids consisting of ursolic,oleanolic, asiatic acids and maslinic	Suppression of the inflammatory cascade through modulation of NFκB signaling pathway	[32, 181, 191]
<i>Salvia miltiorrhiza</i>			Tanshinones	Induces apoptosis for the sub-G1 proportion increase, down-regulation of Bel-2, upregulation of p53 and Bax, and the activation of the caspases-3 and -9 pathways.	[32,156]
<i>Plumbago zeylanica</i>	Roots, stem, flower, and leaves	Petroleum ether, chloroform, ethyl acetate and methanol	1, 4-naphthoquinone	Cell cycle arrest at the G1 phase, apoptosis via the mitochondrial cell death pathway, and enhanced production of reactive oxygen species	[30,182,189]
<i>Psoralea corylifolia</i> (Boh-Gol-Zhee)	Leaves, fruit	Petroleum ether, chloroform, ethanol and aqous	ashoreline, coryfolin and bavachinin	Induction of a long noncoding RNA, RAD51-AS1, which bound to RAD51 mRNA, thereby inhibiting RAD51 protein expression	[30,188,190]
<i>Podophyllum hexandrum</i>		ethyl acetate	odophyllin and podophyllotoxin	-	[30]
<i>Emblica officinalis</i>	Berry, fruit,Leaves, seeds	Ethanol	gallic acid, vitamin C, tannins, flavonoids,	- Prevent mutagenesis and lipid peroxidation in response to carcinogens and reactive oxygen species; Alkylating carcinogens generate DNA mutations through carbon oxidation or conjugation reactions with nucleic acids	[30,184,192]
<i>Stylognecauliflora</i>		Methanol	Oligophenolic compounds SCH 644343 and SCH 644342	-	[157,30]
<i>Elsholtziarugulosa and Thevetia peruviana</i>			Flavonoids (Luteolin and apigenin)	-	[30]
<i>Euphorbia antiqorum</i>	Leaves		Flavonoid and Triterpenoid	-	[30]
<i>Stellerachamaejasme</i>		Methanol	gnidimaacin-pimelea factor stelleramarin, stellerarin,huratoxin, simplex in, sub-toxic, and neochemae-jasmina and B.	Inhibit the proliferation of cancer cell via the down regulation of Smad4-mediated TGF-β signaling pathway	[30,158]
<i>Ginkgo biloba</i>			flavonolglycosides, terpene lactones andginkgolonic acid	Inhibition of proliferation of the cancer cell and triggers apoptosis during the NF-kb/p53 signaling pathway	[30,159]

<i>Glycyrrhiza glabra</i>	Flavonoids(licochalone-A)	Induces apoptosis and cell cycle arrest and improves metabolism through via metabolism through regulation of MIRA.	[30,160]
<i>Gossypium hirsutum</i>	Seeds, roots, and stems	Petroleum ether, methanol and aqueous	Asparagine, gossypol, arginine, resins, [30,185,193]
<i>Ocimum sanctum</i>	Leaves and stem	Methanol, hydroalcoholic, and aqueous	Flavonoids (vicein, cirismaritin, orientin, cirsilienol, isothymusin, apigenin &isothymonin) eugenol, linoleic acid, and rosmarinine acid [30,186,187]
<i>Oldenlandia diffusa</i>	Stem, Leaves	Methanol	oldenlandosides, stigmasterol, beta-sitosterol, flavonoid glycosides,oleanicolic acid, and p-coumaric acid [30,194,195]
<i>Aronia melanocarpa</i>	Fruit		phenolic ingredients, chiefly anthocyanin [30]
<i>Hydrastis Canadensis</i>	Rhizome/Root		Alkaloid (Berberine and hydrastine) berberine, isotetrandrine, berbamine, magnoflorine, columbamine, oxycanthine, chelidone acid and palmatine [161,30]
<i>Berberis vulgaris</i>	Root		- [30]
<i>Aloe vera</i>	Leaves	Diethylether, Ethanol, Water	alkaloids, flavonoids, saponin, phenol, glycosides, and tannins [30,162,163]
<i>Andrographis paniculata</i>	Whole plant	Ethanol	diterpenoids, flavonoids and polyphenols [30,164]
<i>Bauhinia variegata</i>	Root, Stem bark, Flower bud	Ethanol	alkaloids, tannins, steroids, anthraquinones, coumarin, saponins, cardiac glycosides, flavonoids, [30,165,166]
<i>Rheum emodi</i>			physcion, anthraquinone (rhein, chrysophanol, aloce-emodin, emodin, and their glycosides) and stilbene (picetannol, resveratrol and their glycosides [30,167,168]
<i>Ballota nigra</i>	Root and stem	Ethanol, chloroform, and ethyl acetate	Ladanein, phenylpropanoid glycosides, diterpenes, flavonoids, oils and beta-ines [30,169,196]
<i>Melastomadecemfidum</i>	Leaf	Methanol	Naringenin, flavonoids, tannins, phenylpropanoids, organic acids, terpenoids, and steroids [30, 197]
<i>Swietenia macrophylla</i>	Stem		3-hydroxy caruilignan [30]
<i>Magnolia officinalis</i>	bark, leaves, and cones		Lignin (Honokiol) [30]
<i>Blumea balsamifera (sambongis)</i>	Leaves		ketones,terpenoids, phenols, fatty acids, alcohol, ethers, aldehydes, pyridines, furans, and alkanes) and non-volatile (flavonoids, flavanones, and chalcones) [170,30,171]

<i>Panax Ginseng</i>	Root, Rhizomes, Leaves, Stem	Ethanol and methanol	Ginsenosides	Inhibiting cancer cell proliferation and promoting cancer cell apoptosis	[30,198,199]
<i>Astragalus membranaceus</i>	Root	Ethanol	Swainsonine, saponins, flavonoids and polysaccharides	Down-regulate MTIG through daidzein to promote ferroptosis of HCC cells and improve prognosis	[30,200,201]
<i>Betula utilis</i>	stem bark	Methanol	Betulin, triterpenoids, phenolics, and flavonoids	Activation of extrinsic apoptosis pathway via up regulation of DR4, DR5 and PARP cleavage in MCF-7 cells over non-tumorigenic MCF-10A cells	[30,202,203]
<i>Picrorrhizakurroa (Kutki)</i>	Rhizomes, root	Methanolic, alcohol and aqueous	icosteroids-I, II and III and kutkoside	Prevention of biochemical changes in the liver and serum of galactosamine-intoxicated	[30,204]
<i>Foeniculum vulgare</i>	Seed	Methanolic	proteins, cardiac glycosides, saponins, trace elements, flavonoids, sterols, triterpenes, coumarins, volatile oils, and vitamins	—	[30,172]
<i>Achyrocline satureioides (Dc.) Lam,</i>		Methanolic	Flavonoids, chalcone, caffeoyl derivatives	—	[173,174]
<i>Lithraea molleoides (Vell.) Engl.,</i>	fruit, leaves and aerial parts	Alcohol	limonene, α -pinene, β -pinene, α -terpineol, myrcene, sabinene hydrate, 4-terpineol, camphene, and Δ -3-carene	—	[173,175]
<i>Schinus molle L.,</i>	trees and shrubs		Limonene, α -Phellandrene, Camphene, α -Pinene, β -Pinene, β -Myrcene and Caryophyllene	—	[173,176]
<i>Cryptolepis sanguinolenta</i>	Leaves, Seed	Methanol	Alkaloid (Cryptolepine), tannins, and flavones	—	[177,178]

curative potential due to the presence of phytochemicals [30–35]. Plants and herbs have been reported to be used to treat cancer and other diseases using leaf, root, fruit and bark extracts of the selected plants when investigated for cytotoxic properties on HepG2 cells [35].

Phytochemicals in the Treatment of HCC

Plants are vast sources of secondary metabolites that are shown to be potent chemotherapeutic that is used in treating different diseases including metabolic, chronic cardiovascular, neoplastic, and neurodegenerative. Although there is a hopeful indication, but many phytochemicals lack the appropriate solid scientific accuracy because of the difficulty in translating them. This is owing to high cost, expertise, and ethical issues among others complicating the successful clinical translation.

The dietary phytochemicals, such as resveratrol, curcumin, quercetin, N-trans-feruloyl octopamine, emodin, silybin, lycopene, phloretin and caffeine show anti-cancer properties against HCC. The cancer hallmarks are the key molecular targets of the phytochemicals to halt the pathogenesis of HCC [34]. Human health is improved or maintained with the therapeutic potential of several plant-derived compounds against various diseases, including HCC [35]. The delay in tumour progression, increased survival and life quality and improved quality of life are due to the synergistic actions of various medicinal plants and chemotherapy [30]. Many concoctions with different combinations of herbal plants have demonstrated improvement in liver function and promoted liver regeneration as well as prevented liver damage and have also been used to treat hepatitis, liver fibrosis and HCC Table 2 [32]. Plants have been shown to have a diversity of secondary metabolites such as phenolic (e.g., flavonoids, coumarins, phenolic acids, quinones, lignans, tannins, stilbenes), nitrogen-containing compounds (e.g., betalains, amines, alkaloids), terpenoids (e.g. carotenoids), vitamins, and many endogenous metabolites whereas phenolic acids and Flavonoid are important groups of secondary metabolites competent of promoting anti-ageing, scavenging free superoxide radicals, reducing the risk of cancer and advance plant flavour, colouration and aroma while Phenolic compounds serve as defensive agents against invading plant pathogens. Therefore, flavonoids and Phenolic are reported to demonstrate anti-depressant, cytotoxic, anti-

ulcerative, antioxidant, anti-cancer, anti-inflammatory and anti-depressant activities. Antioxidant activity is a fundamental property of most of bioactive compounds with anti-carcinogenic and anti-ageing properties [30,32,36,35].

Herbal medicines are still being extensively used universally, but there persists a huge lag in the reception of their strength by the medical and scientific community owing to the deficient scientific evidence. Advance studies into the compounds are much desirable to assist in closing the current gap between clinical application and pre-clinical research. Plants are adjudged to be rich in secondary metabolites, effective chemotherapeutic and/or chemo-preventive agents used for several chronic metabolic, neoplastic, metabolic, neurodegenerative, and cardiovascular diseases.

Plant-derived products would continue to provide broad options in efficiently treating HCC since these products are essentially abundant in nature, more so they are assumed to lead to effects that are fewer or less toxic side. The medical field, in particular, cancer study is continuously evolving, and futuristically, there is a need for the exploration to understand the properties and potentials of phytochemicals in HCC and other cancer types. Several natural compounds derived from plant sources have been evaluated for their effectiveness as potential treatment options against hepatocellular carcinoma (Table 1).

In-silico analysis of plant-based phytochemicals against HCC

Following rigorous analysis, a few pharmacologically active elements from plants can be used as the most promising therapeutic options for HCC. Chemicals produced by plants have demonstrated anticancer potentials for the treatment of a range of cancers [205]. Numerous plant-derived chemicals have been introduced and used as chemotherapeutic medicines, according to studies [205]. *In silico* methods such as pharmacokinetics, pharmacogenomics, molecular docking, and molecular dynamics simulations are commonly used for the mechanistic prediction of plant-based phytochemicals and target proteins. The predicting properties include absorption, distribution, metabolism excretion and (ADMET) as well as the drug-likeness or drug-ability (Lipinski's rule of five), which is very germane in the prediction of the drug properties of plant-based phytochemicals. To examine the ligands' capacity to bind to target

binding sites, molecular docking and molecular dynamics simulation techniques define the atomic-level interactions between plant phytochemicals as ligand molecules and various proteins as receptor or target molecules [206–209]. The molecular interaction of plant-based phytochemicals with several target proteins (such as Bcl2-associated X (BAX) proteins, glycolytic enzymes, caspase enzymes, human placenta aromatase, poly ADP-ribose polymerase (PARP), phosphoinositide-3-kinase, tumour necrosis factor alpha (TNF- α), epidermal growth factor receptor tyrosine kinase (EGFR), Human placental aromatase cytochrome P450 (CYP19A1), Cyclin D and p53) to revealed mechanism of HCC treatment by numerous plants have been reported [210 – 213]. Despite recent developments in medicines, technology, and screening techniques, HCC still has a low rate of survival and is the deadliest malignancy. It is more likely that ligand molecules identified through in-silico research, such as molecular docking, will advance to the next stage of the drug development process.

Conclusion

Despite the progressions in HCC incidence and prevalence, diagnosis and therapeutics, scientists have not discovered or developed lead compounds for HCC without major side effects. Consequently, advances in alternative therapies are desirable in order to overcome such setbacks. Over 50% of the already existing medicines or natural compounds are derived from plants. Hence, there is a need to offer an immeasurable source for diverse chemical structures with anticancer properties to sustain the drug discovery process. The recent growing applications of molecular biological techniques and combinatorial chemistry approaches provide access to novel natural compounds with anticancer properties for future drug development and discovery.

Medicinal plants are currently used in treating HCC due to their several phytochemical components and high antioxidant and anti-inflammatory properties inhibiting human HepG2 cell propagation. Additionally, these plants with their phytochemicals belong to different classes such as proteins, cardiac glycosides, saponins, trace elements, flavonoids, Phenolic, tannins, sugar, starch sterols, triterpenes, coumarins, volatile oils, and vitamins have shown the ability to affect several signalling pathways, signifying their anti-

tumour potential. Therefore, there might be a superior option for preventing the progression or propagation of HCC.

Future Perspective

Several medicinal plants have shown insignificant side effects even with high safety profile and cost-effectiveness than some synthetic medicines and therapies if taken proportionally. Thus, apart from showing possible synergistic anticancer effects, they can also protect patients from the potential side effects of chemotherapies. However, a few shortcomings of this therapy still need to be addressed. The phytoconstituents or phytochemical low bioavailability, instability, and hydrophobicity hinder the practical clinical application of anticancer therapy to attain its full potential. Functionalized nanomaterials, micelles, liposomes, and phospholipid complexes would address these shortcomings or issues encountered. Therefore, further study must be done to understand their chemical kinetics and dynamic action to understand their therapeutic strategy better and determine the appropriate dose required for effective HCC treatment. Thus, the recent review summarized those medicinal plants that have phytochemicals with feasible natural product sources and promising candidates to be used as an adjuvant in treating HCC. These molecules can be developed into a novel anticancer molecule once their complete apoptotic action has been studied clinically.

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References

1. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. *CA Cancer J Clin* 61(2): 69-90. <https://doi.org/10.3322/caac.20107>.
2. Zhang C, Jia, X, Bao J, Chen S, Wang K, Zhang Y, He C (2015) Polyphyllin VII induces apoptosis in HepG2 cells through ROS-mediated mitochondrial dysfunction and MAPK pathways. *BMC complementary and alternative medicine* 16(1): 1-12. <https://doi.org/10.1186/s12906-016-1036-x>
3. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. *cell* 144(5): 646-674. <https://doi.org/10.1016/j.cell.2011.02.013>.
4. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. (2021) Global cancer statistics 2020: GLOBOCAN estimates of incidence

- and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 71: 209-249. <https://doi.org/10.3322/caac.21660>
5. Dodia S, Annappa B, Mahesh PA (2022) Recent advancements in deep learning based lung cancer detection: A systematic review. *Engineering Applications of Artificial Intelligence* 116: 105490. <https://doi.org/10.1016/j.engappai.2022.105490>
 6. Omara T, Odero MP, Obakiro SB (2022) Medicinal plants used for treating cancer in Kenya: an ethnopharmacological overview. *Bulletin of the National Research Centre*, 46(1): 1-33. <https://doi.org/10.1186/s42269-022-00840-x>
 7. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Bray F (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International journal of cancer*, 136(5): E359-E386. <https://doi.org/10.1002/ijc.29210>
 8. Zhang Z, Zhang C, Ding Y, Zhao Q, Yang L, Ling J, Zhang Y (2014) The activation of p38 and JNK by ROS, contribute to OLO-2-mediated intrinsic apoptosis in human hepatocellular carcinoma cells. *Food and Chemical toxicology* 63: 38-47. <https://doi.org/10.1016/j.fct.2013.10.043>
 9. Ouhtit A, Gaur RL, Abdraboh M, Ireland SK, Rao PN, Raj SG, Raj MH (2013) Simultaneous inhibition of cell-cycle, proliferation, survival, metastatic pathways and induction of apoptosis in breast cancer cells by a phytochemical super-cocktail: genes that underpin its mode of action. *Journal of Cancer* 4(9): 703. doi:10.7150/jca.7235.
 10. Xia Jiang, Jing Tan, Jingsong Li, Saul Kivimäe, Xiaojing Yang, Li Zhuang, Puay Leng Lee, Mark TW Chan, Lawrence W Stanton, Edison T Liu, Benjamin NR Cheyette, Qiang Yu (2008) DACT3 Is an Epigenetic Regulator of Wnt/ β -Catenin Signaling in Colorectal Cancer and Is a Therapeutic Target of Histone Modifications. *Cancer Cell* 13(6): 529-541. <https://doi.org/10.1016/j.ccr.2008.04.019>
 11. Khazaei S, Ramachandran V, Esa NM, Etemad A, Moradipour S, Ismail P (2017) Flower extract of *Allium atroviolaceum* triggered apoptosis, activated caspase-3 and down-regulated antiapoptotic Bcl-2 gene in HeLa cancer cell line. *Biomedicine & Pharmacotherapy* 89: 1216-1226. <https://doi.org/10.1016/j.biopha.2017.02.082>
 12. Dehelean CA, Marcovici I, Soica C, Mioc M, Coricovac D, Iurciuc S, Pinzaru I (2021) Plant-derived anticancer compounds as new perspectives in drug discovery and alternative therapy. *Molecules* 26(4): 1109. <https://doi.org/10.3390/molecules26041109>
 13. Mattiuzzi C, Lippi G (2019) Current cancer epidemiology. *Journal of epidemiology and global health* 9(4): 217. doi:10.2991/jegh.k.191008.001.
 14. Heimbach JK, Kulik LM, Finn RS, Sirlin CB, Abecasis MM, Roberts LR, Marrero JA (2018) AASLD guidelines for the treatment of hepatocellular carcinoma. *Hepatology* 67(1): 358-380. <https://doi.org/10.1002/hep.29086>
 15. Chiang CJ, Yang YW, You SL, Lai MS, Chen CJ (2013) Thirty-year outcomes of the national hepatitis B immunization program in Taiwan. *Jama* 310(9): 974-976. doi:10.1001/jama.2013.276701
 16. Suresh D, Srinivas AN, Kumar DP (2020) Etiology of hepatocellular carcinoma: special focus on fatty liver disease. *Frontiers in Oncology* 10: 601710. <https://doi.org/10.3389/fonc.2020.601710>
 17. Kew KS, Neivashini M, Ooi XC, Nabila P, Khan NH (2018) Qualitative study on the phytochemical constituents of the flower buds of *Bauhinia variegata*. *J. Pharma. Res* 2(1): 47-51. DOI:10.18689/mjpr-1000108
 18. Bajaj M, Blundell TL, Pitts JE, Wood SP, Tatnell MA, Falkmer S, Emdin SO, Gowan LK, Crow H, Schwabe C, Wollmer A, Strassburger W (1983) Dogfish insulin. *European Journal of Biochemistry* 135: 535-542. <https://doi.org/10.1111/j.1432-1033.1983.tb07685.x>
 19. Shen HM, Ong CN (1996) Mutations of the p53 tumor suppressor gene and ras oncogenes in aflatoxin hepatocarcinogenesis. *Mutation Research/Reviews in Genetic Toxicology* 366(1): 23-44. [https://doi.org/10.1016/S0165-1110\(96\)90005-6](https://doi.org/10.1016/S0165-1110(96)90005-6)
 20. Akinyemiju T, Abera S, Ahmed M, Alam N, Alemayohu MA, Allen C (2017) Global Burden of Disease Liver Cancer Collaboration (2017) The burden of primary liver cancer and underlying etiologies from 1990 to 2015 at the global, regional, and national level: results from the global burden of disease study 2015. *JAMA Oncology* 3(12): 1683-1691. doi:10.1001/jamaoncol.2017.3055
 21. Gramenzi A, Caputo F, Biselli M, Kuria F, Loggi E, Andreone P, Bernardi M (2006) Alcoholic liver disease—pathophysiological aspects and risk factors. *Alimentary pharmacology & therapeutics*, 24(8): 1151-1161. <https://doi.org/10.1111/j.1365-2036.2006.03110.x>
 22. Modi SJ, Tiwari A, Kulkarni VM (2021) Reversal of TGF- β -induced epithelial–mesenchymal transition in hepatocellular carcinoma by sorafenib, a VEGFR-2 and Raf kinase inhibitor. *Current Research in Pharmacology and Drug Discovery* 2: 100014. <https://doi.org/10.1016/j.crphar.2021.100014>
 23. Moradpour D, Blum HE (2005) Pathogenesis of hepatocellular carcinoma. *European journal of gastroenterology & hepatology* 17(5): 477-483.
 24. Nault JC, Calderaro J, Di Tommaso L, Balabaud C, Zafrani ES, Bioulac-Sage P, Zucman-Rossi J (2014) Telomerase reverse transcriptase promoter mutation is an early somatic genetic alteration in the transformation of premalignant nodules in hepatocellular carcinoma on cirrhosis. *Hepatology* 60(6): 1983-1992. <https://doi.org/10.1002/hep.27372>
 25. Zhang X, Guan L, Tian H, Zeng Z, Chen J, Huang D, Li Y (2021) Risk factors and prevention of viral hepatitis-related hepatocellular carcinoma. *Frontiers in oncology* 11: 686962. <https://doi.org/10.3389/fonc.2021.686962>
 26. Ezekwesili-Ofili JO, Okaka ANC (2019) Herbal medicines in African traditional medicine, herbal medicine. Philip F. Builders, Intech Open, DOI, 10. <http://dx.doi.org/10.5772/intechopen.80348>
 27. Galabuzi C, Agea J, Fungo B, Kamoga R (2010) Traditional medicine as an alternative form of health care system: a preliminary case study of Nangabo sub-county, central Uganda. *African journal of traditional,*

- complementary, and alternative medicines 7(1). <https://doi.org/10.1007/s13593-018-0533-3>
28. Geck MS, Cristians S, Berger-Gonzalez M, Casu L, Heinrich M, Leonti M (2020) Traditional herbal medicine in Mesoamerica: toward its evidence base for improving universal health coverage. *Frontiers in pharmacology* 11: 1160. <https://doi.org/10.1038/s41591-022-01744-2>.
 29. Sun S, Wang Y, Wu A, Ding Z, Liu X (2019) Influence factors of the pharmacokinetics of herbal resourced compounds in clinical practice. *Evidence-Based Complementary and Alternative Medicine* <https://doi.org/10.1155/2019/1983780>
 30. Mathew S, Faheem M, Suhail M, Fatima K, Archunan G, Begum N, Qadri I (2016) Updates on traditional medicinal plants for hepatocellular carcinoma. *Pharmaceutical Journals* 8(3). DOI:10.5530/pj.2016.3.5.
 31. Nasreen S, Safeer S, Dar KK, Andleeb S, Ejaz M, Khan MA, Ali S (2018) Etiology of hepatocellular carcinoma and treatment through medicinal plants: a comprehensive review. *Oriental Pharmacy and Experimental Medicine*, 18(3): 187-197. DOI:10.1007/s13596-018-330-1
 32. Rawat D, Shrivastava S, Naik RA, Chhonker SK, Mehrotra A, Koiri RK (2018) An overview of natural plant products in the treatment of hepatocellular carcinoma. *Anti-Cancer Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Anti-Cancer Agents)*, 18(13): 1838-1859. DOI: 10.2174/1871520618666180604085612
 33. Tibiri A, Boria S, Traoré TK, Ouédraogo N, Nikiéma A, Ganaba S, Carraz M (2020) Countrywide survey of plants used for liver disease management by traditional healers in Burkina Faso. *Frontiers in Pharmacology* 11: 563751. <https://doi.org/10.3389/fphar.2020.563751>
 34. Rodriguez S, Skeet K, Mehmetoglu-Gurbuz T, Goldfarb M, Karri S, Rocha J, Subramani R (2021) Phytochemicals as an alternative or integrative option, in conjunction with conventional treatments for hepatocellular carcinoma. *Cancers* 13(22): 5753. doi: 10.3390/cancers13225753
 35. Thusyanthan J, Wickramaratne NS, Senathilake KS, Rajagopalan U, Tennekoon KH, Thabrew I, Samarakoon SR (2022) Cytotoxicity against Human Hepatocellular Carcinoma (HepG2) Cells and Anti-Oxidant Activity of Selected Endemic or Medicinal Plants in Sri Lanka. *Advances in Pharmacological and Pharmaceutical Sciences* doi: 10.1155/2022/6407688.
 36. Sagbo JJ, Otang-Mbeng W (2021) Plants used for the traditional management of cancer in the eastern cape province of south africa: A review of ethnobotanical surveys, ethnopharmacological studies and active phytochemicals. *Molecules* 26(15): 4639. doi: 10.3390/molecules26154639.
 37. Perumal A, AlSalhi MS, Kanakarajan S, Devanesan S, Selvaraj R, Tamizhazhagan V (2021) Phytochemical evaluation and anticancer activity of rambutan (*Nephelium lappaceum*) fruit endocarp extracts against human hepatocellular carcinoma (HepG-2) cells. *Saudi Journal of Biological Sciences*, 28(3): 1816-1825. <https://doi.org/10.1016/j.sjbs.2020.12.027>.
 38. Okah Reminus, Walter Cornelius (2019) Phytochemical analysis of moringa oleifera (leaves and flowers) and the functional group.; *GSJ* 7(6).
 39. Dwivedi MK, Sonter S, Mishra S, Patel DK, Singh PK (2020) Antioxidant, antibacterial activity, and phytochemical characterization of *Carica papaya* flowers. *Beni-Suef University Journal of Basic and Applied Sciences* 9(1): 1-11. <https://doi.org/10.1186/s43088-020-00048-w>
 40. Ali M, Ibrahim IS (2019) Phytochemical screening and proximate analysis of garlic (*Allium sativum*). *Inorg Chem* 4: 478-482. <http://dx.doi.org/10.32474/AO-ICS.2019.04.000180>
 41. Zafar H, Rashid MU, Sarvepalli D, Khan MM, Mandzhieva B, Jain AG, Ahmad S (2020) Emerging Roles of Phytochemicals in Hepatocellular Carcinoma. In *Phytochemicals Targeting Tumor Microenvironment in Gastrointestinal Cancers*. Springer, Cham 287-302 DOI:10.1007/978-3-030-48405-7_13
 42. Doreddula SK, Bonam SR, Gaddam DP, Desu BSR, Ramarao N, Pandey V (2014) Phytochemical analysis, antioxidant, antistress, and nootropic activities of aqueous and methanolic seed extracts of ladies finger (*Abelmoschus esculentus* L.) in mice. *The Scientific World Journal* doi: 10.1155/2014/519848.
 43. Chaemsawang W, Prasongchean W, Papadopoulos KI, Ritthidej G, Sukrong S, Wattanaarsakit P (2019) The effect of okra (*Abelmoschus esculentus* (L.) Moench) seed extract on human cancer cell lines delivered in its native form and loaded in polymeric micelles. *International Journal of Biomaterials* 2019. doi: 10.1155/2019/9404383.
 44. Adomèniènè A, Venskutonis PR (2022) *Dioscorea* spp.: Comprehensive Review of Antioxidant Properties and Their Relation to Phytochemicals and Health Benefits. *Molecules* 27(8): 2530. doi: 10.3390/molecules27082530
 45. Kerdput V, Nilbu-Nga C, Kaewnoonual N, Itharat A, Pongsawat S, Pradidarcheep W (2021) Therapeutic efficacy of a *Dioscorea* membranacea extract in a rat model of hepatocellular carcinoma: Histopathological aspects. *Journal of traditional and complementary medicine* 11(5): 400-408. doi: 10.1016/j.jtcme.2021.02.001
 46. Talari M, Seydi E, Salimi A, Mohsenifar Z, Kamalinejad M, Pourahmad J (2014) *Dracocephalum*: novel anticancer plant acting on liver cancer cell mitochondria. *BioMed research international* 2014. <https://doi.org/10.1155/2014/892170>
 47. Kamali M, Khosroyar S, Kamali H, Sani TA, Mohammadi A (2016) Phytochemical screening and evaluation of antioxidant activities of *Dracocephalum kotschyi* and determination of its luteolin content. *Avicenna Journal of Phytomedicine* 6(4): 425. PMID: 27516983; PMCID: PMC4967838.
 48. Aremu AO, Masondo NA, Gruz J, Doležal K, Van Staden J (2019) Potential of smoke-water and one of its active compounds (karrikinolide, KAR1) on the phytochemical and antioxidant activity of *Eucomis autumnalis*. *Antioxidants*, 8(12): 611. doi: 10.3390/antiox8120611
 49. Kaewnoonual N, Itharat A, Pongsawat S, Nilbu-Nga C, Kerdput V, Pradidarcheep W (2020) Anti-angiogenic and anti-proliferative effects of *Benja-ummarit* extract in rats with hepatocellular carcinoma. *Biomedical Reports*, 12(3): 109-120. doi: 10.3892/br.2020.1272
 50. Qiang Y, Zhang XX, Liu H, Xu YR (2014) Chemical constituents of *Alisma orientalis*. *Chemistry of Natural*

- Compounds 49(6): 1143-1145. <https://doi.org/10.1007/s10600-014-0844-9>
51. Biswas G, Nandi S, Kuila D, Acharya K (2017) A comprehensive review on food and medicinal prospects of *Astraeus hygrometricus*. *Pharmacognosy Journal* 9(6). DOI:10.5530/pj.2017.6.125
 52. Tian-Shung WU, Yu-Yi CHAN, Yann-Lii LEU (2000) The Constituents of the Root and Stem of *Aristolochia cucurbitifolia* Hayata and Their Biological Activity. *Chem. Pharm. Bull.* 48(7): 1006—1009. doi: 10.1248/cpb.48.1006
 53. Ruffa MJ, Ferraro G, Wagner ML, Calcagno ML, Campos RH, Cavallaro L (2002) Cytotoxic effect of Argentine medicinal plant extracts on human hepatocellular carcinoma cell line. *Journal of Ethnopharmacology* 79(3): 335-339. [https://doi.org/10.1016/S0378-8741\(01\)00400-7](https://doi.org/10.1016/S0378-8741(01)00400-7)
 54. Shen BB, Yang YP, Yasamin S, Liang NA, Su W, Chen SH, Wang W (2018) Analysis of the Phytochemistry and Bioactivity of the Genus *Polygonum* of Polygonaceae. *Digital Chinese Medicine* 1(1): 19-36. DOI:10.1016/S2589-3777(19)30005-9
 55. Soyngbe OS, Mongalo NI, Makhafola TJ (2018) In vitro antibacterial and cytotoxic activity of leaf extracts of *Centella asiatica* (L.) Urb, *Warburgiasalutaris* (Bertol. F.) Chiov and *Curtisia dentata* (Burm. F.) CA Smedicinal plants used in South Africa. *BMC complementary and alternative medicine* 18(1): 1-10. doi: 10.1186/s12906-018-2378-3
 56. Kemegne GA, Bettache N, Nyegue MA, Etoa FX, Menut C (2020) Cytotoxic activities of *Psidium guajava* and *Mangifera indica* plant extracts on human healthy skin fibroblasts and human hepatocellular carcinoma. *Issues in Biological Sciences and Pharmaceutical Research*, 8(4): 58-64. DOI: 10.15739/ibspr.20.007.
 57. Serunjogi D (2020) Active phytochemicals present in the Guava Tree (*Psidium Guajava*) leaf Extracts that grow in Uganda. *Student's Journal of Health Research Africa* 1(12): 6-6. DOI: <https://doi.org/10.51168/sjhraf-rica.v1i12.12>.
 58. Afolabi AA, Ashamu EA, Oluranti OI (2020) Ameliorative effect of *Psidium guajava* (L.) leaf aqueous extract on aluminium nitrate-induced liver damage in female Wistar rats. *Clinical Phytoscience* 6(1): 1-4. <https://doi.org/10.1186/s40816-020-00198-5>
 59. Abdul Wahab, Rasheed Ahmad Khera, Rafia Rehman, Ayesha Mushtaq, Aicha Blama Merzaia, Muhammad Waqar Azeem (2016) A review on phytochemistry and medicinal uses of *Taxus wallichiana* L. (*Himalayan Yew*). *IJCBS* 9: 116-120.
 60. Li L, Zhang W, Desikan Seshadri VD, Cao G (2019) Synthesis and characterization of gold nanoparticles from *Marsdeniataenacissima* and its anticancer activity of liver cancer HepG2 cells. *Artificial cells, nanomedicine, and biotechnology* 47(1): 3029-3036. doi 10.1080/21691401.2019.1642902
 61. Mondal M, Saha S, Hossain M, Al Foyjul I, Sarkar C, Hossain S, Kundu SK. (2020) Phytochemical profiling and evaluation of bioactivities of methanolic and ethyl acetate extracts of *Marsdeniataenacissima* leaves. *Journal of Herbs, Spices & Medicinal Plants* 26(4): 405-422. DOI:10.1080/10496475.2020.1748784
 62. Li S, Pei W, Yuan W, Yu D, Song H, Zhang H (2022) Multi-omics joint analysis reveals the mechanism of action of the traditional Chinese medicine *Marsdeniataenacissima* (Roxb.) Moon in the treatment of hepatocellular carcinoma. *Journal of Ethnopharmacology* 293: 115285. <https://doi.org/10.1016/j.jep.2022.115285>.
 63. Janjušević L, Karaman M, Šibul F, Tommonaro G, Iodice C, Jakovljević D, Pejin B (2017) The lignicolous fungus *Trametes versicolor* (L.) Lloyd (1920): a promising natural source of antiradical and AChE inhibitory agents. *Journal of Enzyme Inhibition and Medicinal Chemistry* 32(1): 355-362. doi: 10.1080/14756366.2016.1252759
 64. Chang Y, Zhang M, Jiang Y, Liu Y, Luo H, Hao C, Zhang L (2017) Preclinical and clinical studies of *Coriolus versicolor* polysaccharopeptide as an immunotherapeutic in China. *Discovery Medicine* 23(127): 207-219. PMID: 28595034
 65. Kivrak I, Kivrak S, Karababa E (2020) Assessment of bioactive compounds and antioxidant activity of Turkey tail medicinal mushroom *Trametes versicolor* (Agaricomycetes). *International Journal of Medicinal Mushrooms* 22(6). doi: 10.1615/IntJMedMushrooms.2020035027
 66. Tel-Cayan G, Deveci E, Cayan F, Duru ME (2018) Comparative assessment of phytochemical composition, antioxidant and anticholinesterase activities of two Basidiomycota Truffle Fungi from Turkey. *Marmara Pharmaceutical Journal* 22(1): 59-65. DOI:10.12991/mpj.2018.41
 67. Gomes DCV, de Alencar MVOB, Dos Reis AC, de Lima RMT, de Oliveira Santos JV, da Mata AMOF, Cavalcante AADCM (2019) Antioxidant, anti-inflammatory and cytotoxic/antitumoral bioactives from the phylum Basidiomycota and their possible mechanisms of action. *Biomedicine & Pharmacotherapy*, 112: 108643. DOI: 10.1016/j.biopha.2019.108643
 68. Urech K, Baumgartner S (2015) Chemical constituents of *Viscum album* L.: implications for the pharmaceutical preparation of mistletoe. *Mistletoe: From mythology to evidence-based medicine* 4: 11-23. DOI:10.1159/000375422
 69. Yang P, Jiang Y, Pan Y, Ding X, Rhea P, Ding J, Lee RT (2019) Mistletoe extract Fraxini inhibits the proliferation of liver cancer by down-regulating c-Myc expression. *Scientific reports* 9(1): 1-12. DOI: 10.1038/s41598-019-41444-2
 70. Yang Y, Ju Z, Yang Y, Zhang Y, Yang L, Wang Z (2021) Phytochemical analysis of *Panax* species: a review. *Journal of Ginseng Research* 45(1): 1-21. doi: 10.1016/j.jgr.2019.12.009
 71. Jang SI, Lee YW, Cho CK, Yoo HS, Jang JH (2013) Identification of target genes involved in the antiproliferative effect of enzyme-modified ginseng extract in HepG2 hepatocarcinoma cell. *Evidence-Based Complementary and Alternative Medicine* 2013. doi: 10.1155/2013/502568.
 72. Pan J, Yang C, Jiang Z, Huang J (2019) *Trametesrobiniophila*Murr: a traditional Chinese medicine with potent anti-tumor effects. *Cancer Management and Research* 11: 1541. DOI: 10.2147/CMAR.S193174

73. Muñoz-Castiblanco T, Mejía-Giraldo JC, Puertas-Mejía MA (2020) Trametes genus, a source of chemical compounds with anticancer activity in human osteosarcoma: A systematic review. *Journal of Applied Pharmaceutical Science* 10(10): 121-129. DOI: 10.7324/JAPS.2020.1010014
74. Zhang Y, Wang X, Chen T (2019) Efficacy of Huaier granule in patients with breast cancer. *Clin. Transl. Oncol* 21: 588-595. <https://doi.org/10.1007/s12094-018-1959-4>
75. Chen M, Wu J, Luo Q, Mo S, Lyu Y, Wei Y, Dong J (2016) The anticancer properties of *HerbaEpimedii* and its main bioactive components icaritin and icaridin II. *Nutrients*, 8(9): 563. doi: 10.3390/nu8090563
76. Li M, Shang H, Wang T, Yang SQ, Li L (2021) Huanglian decoction suppresses the growth of hepatocellular carcinoma cells by reducing CCNB1 expression. *World Journal of Gastroenterology* 27(10): 939. doi: 10.3748/wjg.v27.i10.939
77. Li S, Wang P (2014) Phytochemistry of *CamptothecaDecaisne*. *Pharmaceutical Crops* 5(1): 163-172. DOI:10.2174/2210290601405010163
78. He H, Shang XY, Liu WW, Zhang Y, Song SJ (2019) Triterpenes from the fruit of *Camptotheca acuminata* suppress human hepatocellular carcinoma cell proliferation through apoptosis induction. *Natural product research* 33(24): 3527-3532. doi: 10.1080/14786419.2018.1487967
79. Juliana Garcia, Francisca Rodrigues, Maria José Saavedra, Fernando M Nunes, Guilhermina Marques (2022) Bioactive polysaccharides from medicinal mushrooms: A review on their isolation, structural characteristics and antitumor activity, *Food Bioscience*, Vol 49, <https://doi.org/10.1016/j.fbio.2022.101955>.
80. Butnariu M, Quispe C, Herrera-Bravo J, Pentea M, Sarac I, Küşümler AS, Cho WC (2022) *Papaver* Plants: Current Insights on Phytochemical and Nutritional Composition Along with Biotechnological Applications. *Oxidative Medicine and Cellular Longevity* 2022. <https://doi.org/10.1155/2022/2041769>.
81. Tanmoy G, Arijit M, Tanushree S, Jagadish S, Kumar MT (2014) Pharmacological actions and phytoconstituents of *Amaranthus spinosus* Linn: a review. *Int J Pharmacogn Phytochem Res* 6: 405-413.
82. Liu J, Cao C, Ding P, Jiang J (2015) Anticancer activity and mechanism of apoptosis induced by *Amaranthus spinosus* L. extract in HepG2 cells. *Chinese Pharmacological Bulletin* 1558-1561. DOI: 10.3969/j.issn.1001-1978.2015.11.016
83. Huang X, Kojima-Yuasa A, Norikura T, Kennedy DO, Hasuma T, Matsui-Yuasa I (2007) Mechanism of the anti-cancer activity of *Zizyphus jujuba* in HepG2 cells. *The American journal of Chinese medicine* 35(03): 517-532. doi: 10.1142/S0192415X0700503X
84. Zhang C, Jia X, Bao J, Chen S, Wang K, Zhang Y, He C (2015) Polyphyllin VII induces apoptosis in HepG2 cells through ROS-mediated mitochondrial dysfunction and MAPK pathways. *BMC complementary and alternative medicine* 16(1): 1-12. DOI: <https://doi.org/10.1186/s12906-016-1036-x>.
85. Choi P, Park JY, Kim T, Park SH, Kim HK, Kang KS, Ham J (2015) Improved anticancer effect of ginseng extract by microwave-assisted processing through the generation of ginsenosides Rg3, Rg5 and Rk1. *Journal of Functional Foods* 14: 613-622. DOI:10.1016/j.jff.2015.02.038
86. Alias NZ, Kamisah N, Ishak M (2014) Chemical Constituents and Bioactivity Studies of *Ardisia elliptica*. In *The Open Conference Proceedings Journal* 5(1). DOI: 10.2174/2210289201405020001
87. Ning-Ning SONG, Lei-Min YANG, Zhang MJ, Ren-Feng AN, Wei LIU, Huang XF (2021) Triterpenoid saponins and phenylpropanoid glycoside from the roots of *Ardisia crenata* and their cytotoxic activities. *Chinese Journal of Natural Medicines* 19(1): 63-69. Corpus ID: 219144045.
88. Lou L, Ye W, Chen Y, Wu S, Jin L, He J, Wang J (2012) Ardisiosilolide inhibits survival, invasion and metastasis of human hepatocellular carcinoma cells. *Phytomedicine*, 19(7): 603-608. DOI: 10.1016/j.phymed.2012.01.003
89. Kaska A, Deniz N, Çiçek M, Mammadov R (2020) The screening of *Digitalis ferruginea* L. subsp. *ferruginea* for toxic capacities, phenolic constituents, antioxidant properties, mineral elements and proximate analysis. *Food Science and Technology* 41: 505-512. <https://doi.org/10.1590/fst.08620>
90. Reddy D, Kumavath R, Ghosh P, Barh D (2019) Lanatoside C induces G2/M cell cycle arrest and suppresses cancer cell growth by attenuating MAPK, Wnt, JAK-STAT, and PI3K/AKT/mTOR signaling pathways. *Biomolecules* 9(12): 792. doi: 10.3390/biom9120792
91. Kwon J, Park JE, Lee JS, Lee JH, Hwang H, Jung SH, Jang DS (2021) Chemical Constituents of the Leaves of *Diospyros kaki* (Persimmon). *Plants* 10(10): 2032. <https://doi.org/10.3390/plants10102032>
92. Kim HS, Suh JS, Jang YK, Ahn SH, Raja G, Kim JC, Kim TJ (2020) Anti-cancer potential of persimmon (*Diospyros kaki*) leaves via the PDGFR-Rac-JNK pathway. *Scientific Reports* 10(1): 1-13. doi: 10.1038/s41598-020-75140-3
93. Gawron-Gzella A, Witkowska-Banaszczak E, Bylka W, Dudek-Makuch M, Odwrot A, Skrodzka N (2016) Chemical composition, antioxidant and antimicrobial activities of *Sanguisorba officinalis* L. extracts. *Pharmaceutical Chemistry Journal* 50(4): 244-249. DOI:10.1007/s11094-016-1431-0
94. Jiang N, Li H, Sun Y, Zeng J, Yang F, Kantawong F, Wu J (2021) Network Pharmacology and Pharmacological Evaluation Reveals the Mechanism of the *Sanguisorba Officinalis* in Suppressing Hepatocellular Carcinoma. *Frontiers in pharmacology* 12: 618522. <https://doi.org/10.3389/fphar.2021.618522>
95. Zhang X, Guan L, Tian H, Zeng Z, Chen J, Huang D, Li Y (2021) Risk factors and prevention of viral hepatitis-related hepatocellular carcinoma. *Frontiers in oncology*, 11: 686962. doi: 10.3389/fonc.2021.686962
96. Guo H, Chen Y, Wang J, Ma H, Liu Y (2022) A Critical Review: Anti-Cancer Effects of *Brucea Javanica* and the Mechanisms. *Pharmacological Research-Modern Chinese Medicine* 100133. <https://doi.org/10.1016/j.prmcm.2022.100133>.
97. Kim BM, Kim GT, Kim EJ, Lim EG, Kim SY, Kim YM (2016) Extract from *Artemisia annua* Linné induces apoptosis through the mitochondrial signaling pathway

- in HepG2 Cells. *Journal of the Korean Society of Food Science and Nutrition* 45(12): 1708-1716. <http://dx.doi.org/10.3746/jkfn.2016.45.12.1708>.
98. Mamatova AS, Korona-Głowniak I, Skalicka-Woźniak K, Józefczyk A, Wojtanowski KK, Baj T, Malm A (2019) Phytochemical composition of wormwood (*Artemisia gmelinii*) extracts in respect of their antimicrobial activity. *BMC Complementary and Alternative Medicine* 19(1): 1-8. doi: 10.1186/s12906-019-2719-x
 99. Picerno P, Sansone F, Mencherini T, Prota L, Aquino RP, Rastrelli L, Lauro MR (2011) Citrus bergamia juice: phytochemical and technological studies. *Natural Product Communications*, 6(7) DOI:10.1177/1934578X1100600707
 100. Ferlazzo N, Cirmi S, Russo M, Trapasso E, Ursino MR, Lombardo GE, Navarra M (2016) NF- κ B mediates the antiproliferative and proapoptotic effects of bergamot juice in HepG2 cells. *Life sciences* 146: 81-91. DOI:10.1016/j.lfs.2015.12.040
 101. Lu JZ, Ye D, Ma BL (2021) Constituents, pharmacokinetics, and pharmacology of Gegen-Qinlian decoction. *Frontiers in Pharmacology* 12, <https://doi.org/10.3389/fphar.2021.668418>
 102. Pan TL, Wang PW, Leu YL, Wu TH, Wu TS (2012) Inhibitory effects of *Scutellaria baicalensis* extract on hepatic stellate cells through inducing G2/M cell cycle arrest and activating ERK-dependent apoptosis via Bax and caspase pathway. *Journal of Ethnopharmacology* 139(3): 829-837. DOI: 10.1016/j.jep.2011.12.028
 103. Rui Jing, Hua-Qiang Li, Chang-Ling Hu, Yi-Ping Jiang, Lu-Ping Qin, Cheng-Jian Zheng (2016) Phytochemical and Pharmacological Profiles of Three Fagopyrum Buckwheats. *Int. J. Mol. Sci* 17: 589; doi:10.3390/ijms17040589
 104. Peng W, Hu C, Shu Z, Han T, Qin L, Zheng C (2015) Antitumor activity of tatariside F isolated from roots of *Fagopyrum tataricum* (L.) Gaertn against H22 hepatocellular carcinoma via up-regulation of p53. *Phytomedicine* 22(7-8): 730-736. DOI: 10.1016/j.phymed.2015.05.003
 105. Elshamy AI, Mohamed TA, Marzouk MM, Hussien TA, Umeyama A, Hegazy MEF, Efferth T (2018) Phytochemical constituents and chemosystematic significance of *Pulicaria jaubertii* E. Gamal-Eldin (Asteraceae). *Phytochemistry Letters* 24: 105-109. <https://doi.org/10.1016/j.phytol.2018.01.021>.
 106. Algabr M, Hajj NA, Dunia AMA, Romane A, Wadhaf HA (2016) Screening of Yemeni medicinal plant for antibacterial and antifungal activities. *Advances in Natural and Applied Sciences* 10(8)
 107. Usman A, Sanusi SB, Lawal SM, Musa FM, Auwal HM (2021) Antibacterial and phytochemical screening of *Ziziphus jujuba* (jujube/magarya) leaf extract in Kaduna Metropolis. *Dutse Journal of Pure and Applied Sciences (DUJOPAS)* 7(4b) <https://dx.doi.org/10.4314/dujopas.v7i4b.19>
 108. Cheng-Yu Hung, Kuo-Hsiung Wang, Chi-Chun Huang, Xun Gong, Xue-Jun Ge, Tzen-Yuh Chiang (2008) Isolation and characterization of 11 microsatellite loci from *Camellia sinensis* in Taiwan using PCR-based isolation of microsatellite arrays (PIMA). *Conserv Genet* 9: 779-781 <https://doi.org/10.1007/s10592-007-9391-2>
 109. Falla NM, Demasi S, Caser M, Scariot V (2021) Phytochemical profile and antioxidant properties of Italian green tea, a new high quality niche product. *Horticulturae* 7(5): 91. <https://doi.org/10.3390/horticulturae7050091>.
 110. Rohman A, Arifah FH, Alam G, Rafi M (2020) A review on phytochemical constituents, role on metabolic diseases, and toxicological assessments of underutilized part of *Garcinia mangostana* L. fruit. *Journal of Applied Pharmaceutical Science* 10(7): 127-146. DOI: 10.7324/JAPS.2020.10716
 111. Rizaldy D, Hartati R, Nadhifa T, Fidrianny I (2021) Chemical Compounds and Pharmacological Activities of Mangosteen (*Garcinia mangostana* L.)—Updated Review. *Biointerface Res. Appl. Chem* 12: 2503-2516. <https://doi.org/10.33263/BRIAC122.25032516>.
 112. Wudtiwai B, Pitchakarn P, Banjerdpongchai R (2018) Alpha-mangostin, an active compound in *Garcinia mangostana*, abrogates anoikis-resistance in human hepatocellular carcinoma cells. *Toxicology in Vitro* 53: 222-232. doi: 10.1016/j.tiv.2018.09.003
 113. Meng FC, Wu ZF, Yin ZQ, Lin LG, Wang R, Zhang QW (2018) *Coptidis rhizoma* and its main bioactive components: recent advances in chemical investigation, quality evaluation and pharmacological activity. *Chinese Medicine* 13(1): 1-18. DOI:10.1186/s13020-018-0171-3
 114. Wang J, Wang L, Lou GH, Zeng HR, Hu J, Huang QW, Yang XB (2019) *Coptidis Rhizoma*: a comprehensive review of its traditional uses, botany, phytochemistry, pharmacology and toxicology. *Pharmaceutical Biology* 57(1): 193-225. DOI: 10.1080/13880209.2019.1577466.
 115. Chai FN, Zhang J, Xiang HM, Xu HS, Li YF, Ma WY, Ye XL (2018) Protective effect of Coptisine from *Rhizoma Coptidis* on LPS/D-GalN-induced acute liver failure in mice through up-regulating expression of miR-122. *Biomedicine & Pharmacotherapy* 98: 180-190. doi: 10.1016/j.biopha.2017.11.133
 116. Paudel KR, Panth N (2015) Phytochemical profile and biological activity of *Nelumbo nucifera*. *Evidence-Based Complementary and Alternative Medicine*, <https://doi.org/10.1155/2015/789124>
 117. Rho T, Yoon KD (2017) Chemical constituents of *Nelumbo nucifera* seeds. *Natural Product Sciences* 23(4): 253-257. DOI: <https://doi.org/10.20307/nps.2017.23.4.253>
 118. Poornima P, Quency RS, Padma VV (2013) Neferine induces reactive oxygen species mediated intrinsic pathway of apoptosis in HepG2 cells. *Food Chemistry* 136(2): 659-667. doi: 10.1016/j.foodchem.2012.07.112
 119. Murthy HN, Dalawai D, Dewir YH, Ibrahim A (2020) Phytochemicals and biological activities of *Garcinia morella* (Gaertn.) Desr.: A review. *Molecules* 25(23): 5690. <https://doi.org/10.3390/molecules25235690>.
 120. Mohd Fadzelly Abu Bakar, Nor Ezani Ahmad, Monica Suleiman, Asmah Rahmat, Azizul Isha, (2015) "*Garcinia dulcis* Fruit Extract Induced Cytotoxicity and Apoptosis in HepG2 Liver Cancer Cell Line", *BioMed Research International* 2015(10), Article ID 916902 <https://doi.org/10.1155/2015/916902>
 121. Gao D, Cho CW, Vinh LB, Kim JH, Kim YH, Kang JS (2021) Phytochemical analysis of trifoliolate orange

- during fermentation by HPLC–DAD–ESI–MS/MS coupled with multivariate statistical analysis. *Acta Chromatographica* 33(4): 371-377. DOI:10.1556/1326.2020.00818
122. Basri AM, Taha H, Ahmad N (2017) A review on the pharmacological activities and phytochemicals of *Alpinia officinarum* (Galangal) extracts derived from bioassay-guided fractionation and isolation. *Pharmacognosy Reviews* 11(21): 43. doi: 10.4103/phrev.phrev_55_16
 123. Singh S, Sahoo BC, Kar SK, Sahoo A, Nayak S, Kar B, Sahoo S (2020) Chemical constituents Analysis of *Alpinia galanga* and *Alpinia calcarata*. *Research Journal of Pharmacy and Technology* 13(10): 4735-4739. DOI:10.5958/0974-360X.2020.00834.3
 124. Blondeau D, St-Pierre A, Bourdeau N, Bley J, Lajeunesse A, Desgagné-Penix I (2020) Antimicrobial activity and chemical composition of white birch (*Betula papyrifera* Marshall) bark extracts. *Microbiologyopen* 9(1), doi: 10.1002/mbo3.944
 125. Qi Y, Zhang Q, Zhu H (2019) Huang-Lian Jie-Du decoction: a review on phytochemical, pharmacological and pharmacokinetic investigations. *Chinese Medicine* 14(1): 1-22. <https://doi.org/10.1186/s13020-019-0277-2>
 126. Reshma BC, Gaikwad DK (2010) Phytochemical Constituents of Tit-Berry (*Allophylus Cobbe* (L.) Raeusch). *BIOINFOLET-A Quarterly Journal of Life Sciences* 7(2): 121-122.
 127. Ghagane SC, Puranik SI, Nerli RB, Hiremath MB (2017) Evaluation of in vitro antioxidant and anticancer activity of *Allophylus cobbe* leaf extracts on DU-145 and PC-3 human prostate cancer cell lines. *Cytotechnology* 69(1): 167-177. DOI: 10.1007/s10616-016-0048-1
 128. Devi N, Sangeetha R (2016) *Madhuca longifolia* (Sapotaceae): A review of its phytochemical and pharmacological profile. *Int. J. Pharmacogen. Biosci* 7: 106-114. DOI:10.22376/ijpbs.2016.7.4.b106-114
 129. Hullatti KK, Gopikrishna UV, Kuppast IJ (2011) Phytochemical investigation and diuretic activity of *Cyclepeltata* leaf extracts. *Journal of Advanced Pharmaceutical Technology & Research*, 2(4): 241. DOI: 10.4103/2231-4040.90880
 130. BCV BPS, Bhat PR (2019) Phytochemical, antimicrobial, antioxidant and immunomodulatory studies of leaf extracts of *Cyclepeltata* (Lam.) Hook. f. & Thomson. *GSC Biological and Pharmaceutical Sciences* 9(3): 052-063. DOI: <https://doi.org/10.30574/gscbps.2019.9.3.0228>
 131. Samaranada VA, Wijekumar PJ, Samarakoon DNAW, Perera PK (2021) The phytochemical constituents and pharmacological properties of *Munronia pinnata*: A review. *Int J Herb Med*, 9(4): 85-91.
 132. Teles YC, Souza MSR, Souza MDFVD (2018) Sulphated flavonoids: biosynthesis, structures, and biological activities. *Molecules* 23(2): 480. <https://doi.org/10.3390/molecules23020480>.
 133. Junejo JA, Zaman K, Rudrapal M, Monda I P, Singh KD, Verma VK (2014) Preliminary phytochemical and physicochemical evaluation of *Caralliabrachiata* (Lour.) Merr. leaves. *Journal of applied pharmaceutical science*, 4(12): 123-127. DOI:10.7324/JAPS.2014.41221.
 134. Gull N, Arshad F, Naikoo GA, Hassan IU, Pedram MZ, Ahmad A, Tambuwala MM (2022) Recent Advances in Anticancer Activity of Novel Plant Extracts and Compounds from *Curcuma longa* in Hepatocellular Carcinoma. *Journal of Gastrointestinal Cancer* 1-23. doi: 10.1007/s12029-022-00809-z
 135. Nath LR, Gorantla JN, Thulasidasan AKT, Vijayakurup V, Shah S, Anwer S, Anto RJ (2016) Evaluation of utroside B, a saponin from *Solanum nigrum* Linn, as a promising chemotherapeutic agent against hepatocellular carcinoma. *Scientific reports* 6(1): 1-13. doi: 10.1038/srep36318.
 136. Ahmed HM (2018) Ethnomedicinal, phytochemical and pharmacological investigations of *Perilla frutescens* (L.) Britt. *Molecules* 24(1): 102. doi: <https://doi.org/10.3390%2Fmolecules24010102>
 137. Najm OA, Addnan FH, Mohd-Manzor NF, Elkadi MA, Abdullah WO, Ismail A, Mansur FAF (2021) Identification of Phytochemicals of *Phoenix dactylifera* L. CvAjwa with UHPLC-ESI-QTOF-MS/MS. *International Journal of Fruit Science* 21(1): 848-867 DOI: 10.1080/15538362.2021.1939227.
 138. Siddiqui S, Ahmad R, Khan MA, Upadhyay S, Husain I, & Srivastava AN (2019) Cytostatic and anti-tumor potential of Ajwa date pulp against human hepatocellular carcinoma HepG2 cells. *Scientific reports* 9(1): 1-12. DOI: <https://doi.org/10.1038/s41598-018-36475-0>
 139. Chanda S Ramachandra TV (2019) Phytochemical and pharmacological importance of turmeric (*Curcuma longa*): A review. *Research & Reviews: A Journal of Pharmacology* 9(1): 16-23.
 140. de Lima RMT, Dos Reis AC, de Menezes AAPM, Santos JVDO, Filho JWGDO, Ferreira JRDO, Melo-Cavalcante AADC (2018) Protective and therapeutic potential of ginger (*Zingiber officinale*) extract and [6]-gingerol in cancer: A comprehensive review. *Phytotherapy research* 32(10): 1885-1907. doi: 10.1002/ptr.6134.
 141. Amin A, Hamza AA, Bajbouj K, Ashraf SS, Daoud S (2011) Saffron: a potential candidate for a novel anticancer drug against hepatocellular carcinoma. *Hepatology* 54(3): 857-867. <https://doi.org/10.1002/hep.24433>.
 142. Nigam M, Atanassova M, Mishra AP, Pezzani R, Devkota HP, Plygun S, Sharifi-Rad J (2019) Bioactive compounds and health benefits of *Artemisia* species. *Natural product communications* 14(7), <https://doi.org/10.1177/1934578X19850354>.
 143. Bisht D, Kumar D, Kumar D, Dua K, Chellappan DK (2021) Phytochemistry and pharmacological activity of the genus *artemisia*. *Archives of Pharmacal Research* 44(5): 439-474. doi: <https://doi.org/10.1007%2F12272-021-01328-4>
 144. Raja Madhan, K Selvakumar, Vidya DJ, Krishna PK, Jayaraman K (2012) Isolation and identification of essential phytochemical constituents of *amorphophallus campanulatus* and screening its antimicrobial properties. *Research Journal of Pharmacy and Technology* 5: 219-225.
 145. Srivastava S, Verma D, Srivastava A, Tiwari SS, Dixit B, Singh RS, Rawat AKS (2014) Phytochemical and nutritional evaluation of *Amorphophallus campanulatus* (Roxb.) Blume Corm. *Journal of Nutrition & Food Sciences* 4(3): 1. DOI:10.4172/2155-9600.1000274

146. Ansil PN, Wills PJ, Varun R, Latha MS (2014) Cytotoxic and apoptotic activities of *Amorphophallus campanulatus* tuber extracts against human hepatoma cell line. *Research in Pharmaceutical Sciences* 9(4): 269. PMC4314875
147. Choa John Benson D, Lu Roanne V, Nombrado Mark A, Rayos Garina Kaye R, Invento Chelsea Dae, Castañeda Gerald (2016) Phytochemical screening of *Broussonetia luzonensis* (Moraceae) leaves, *Journal of Chemical and Pharmaceutical Research* 8(2): 335-338
148. Chao WW, Chen SJ, Peng HC, Liao JW, Chou ST (2019) Antioxidant activity of *Graptopetalum paraguayense* E. Walther leaf extract counteracts oxidative stress induced by ethanol and carbon tetrachloride Co-induced hepatotoxicity in rats. *Antioxidants* 8(8): 251. <https://doi.org/10.3390/antiox8080251>
149. Hsu WH, Chang CC, Huang KW, Chen YC, Hsu SL, Wu LC, Huang CYF (2015) Evaluation of the medicinal herb *Graptopetalum paraguayense* as a treatment for liver cancer. *PloSone* 10(4): e0121298. <https://doi.org/10.1371/journal.pone.0121298>.
150. Dalli M, Bekkouch O, Azizi SE, Azghar A, Geyra N, Kim B (2021) *Nigella sativa* L. Phytochemistry and Pharmacological Activities: A Review (2019–2021). *Biomolecules* 12(1): 20. <https://doi.org/10.3390/biom12010020>.
151. Khan SH, Ansari J, Haq AU, Abbas G (2012) Black cumin seeds as phyto-genic product in broiler diets and its effects on performance, blood constituents, immunity and caecal microbial population. *Italian Journal of Animal Science* 11(4) <https://doi.org/10.4081/ijas.2012.e77>.
152. Lee NCW, Carella MA, Papa S, Bubici C (2018) High Expression of Glycolytic Genes in Cirrhosis Correlates With the Risk of Developing Liver Cancer. *Front. Cell Dev. Biol* 6: 138. doi: 10.3389/fcell.2018.00138
153. Kim HJ, Park SY, Lee HM, Seo DI, Kim YM (2015) Antiproliferative effect of the methanol extract from the roots of *Petasites japonicus* on Hep3B hepatocellular carcinoma cells in vitro and in vivo. *Experimental and therapeutic medicine* 9(5): 1791-1796. <https://doi.org/10.3892/etm.2015.2296>.
154. Verma S, Khambhala P, Joshi S, Kothari V, Patel T, Seshadri S (2019) Evaluating the role of dithiolane rich fraction of *Ferula asafoetida* (apiaceae) for its antiproliferative and apoptotic properties: in vitro studies. *Exp. Oncol*, 4141(22): 90-94. <https://doi.org/10.32471/exp-oncology.2312-8852.vol-41-no-2.12989>.
155. Rehman G, Hamayun M, Iqbal A, Khan SA, Khan H, Shehzad A, Lee IJ (2017) Effect of methanolic extract of dandelion roots on cancer cell lines and AMP-activated protein kinase pathway. *Frontiers in pharmacology* 8: 875. <https://doi.org/10.3389/fphar.2017.00875>.
156. Hu S, Chen SM, Li XK, Qin R, Mei ZN (2007) Antitumor effects of chi-shen extract from *Salvia miltiorrhiza* and *Paeoniae radix* on human hepatocellular carcinoma cells. *Acta Pharmacologica Sinica* 28(8): 1215-1223. <https://doi.org/10.1111/j.1745-7254.2007.00606.x>.
157. Hegde VR, Pu H, Patel M, Das PR, Butkiewicz N, Arreaza G, Chan TM (2003) Two antiviral compounds from the plant *Stylognecauliflora* as inhibitors of HCV NS3 protease. *Bioorganic & medicinal chemistry letters* 13(17): 2925-2928. [https://doi.org/10.1016/s0960-894x\(03\)00584-5](https://doi.org/10.1016/s0960-894x(03)00584-5).
158. Pan G, Cheng L, Feng X, Zhu X, Wu G (2017) Ethanol Extract of *Stellerachamaejasme* L. Inhibits Hepatoma Cell Proliferation Through Down-regulation of Smad4-mediated TGF-beta Signaling Pathway. *International Journal of Pharmacology* 13(6): 628-635. <https://doi.org/10.3923/ijp.2017.628.635>.
159. Wang R, Shao X, Yang J, Liu Z, Chew L, Shao Y (2020) Ginkgo biloba extract mechanism inhibits hepatocellular carcinoma through the nuclear factor- κ B/p53 signaling pathway. *Journal of Environmental Pathology, Toxicology and Oncology* 39(2). <https://doi.org/10.1615/jenvironpatholtoxicol-col.2020034510>.
160. Abdel-WahabAHA., Effat H, Mahrous EA, Ali MA, Al-Shafie TA (2021) A licorice roots extract induces apoptosis and cell cycle arrest and improves metabolism via regulating MiRNAs in liver cancer cells. *Nutrition and Cancer* 73(6): 1047-1058. <https://doi.org/10.1080/01635581.2020.1783329>.
161. Burkhart, EP, Zuiderveen GH (2019). Wild Goldenseal (*Hydrastis canadensis*) Rhizome/Root Alkaloid Content in Relation to Colony and Harvest Stage. *Journal of Herbs, Spices & Medicinal Plants* 25(2): 128-140. <https://doi.org/10.1080/10496475.2019.1577322>.
162. Ikpe V, Eze CS, Mbaaji P, Joshua PE (2017) Phytochemical Analysis and Antifungi Activity of Aloe Vera Leaves. *Bio-Research* 15: 974-979. <https://doi.org/10.4314/br.v15i1.188320>.
163. Nalimu F, Oloro J, Kahwa I, Ogwang, PE (2021). Review on the phytochemistry and toxicological profiles of Aloe vera and Aloe ferox. *Future Journal of Pharmaceutical Sciences* 7(1): 1-21. <https://doi.org/10.1186%2Fs43094-021-00296-2>
164. Chao WW, Lin BF (2010) Isolation and identification of bioactive compounds in *Andrographis paniculata* (Chuanxinlian). *Chinese medicine* 5(1): 1-15. <https://doi.org/10.1186/1749-8546-5-17>
165. Shahana S, Nikalje APG, Nikalje G (2017). A brief review on *Bauhinia variegata*: phytochemistry, antidiabetic and antioxidant potential. *Am. J. Pharmtech Res* 7: 186-197.
166. Kew KS, Neivashini M, Ooi XC, Nabila P, Khan NH (2018). Qualitative study on the phytochemical constituents of the flower buds of *Bauhinia variegata*. *J. Pharma. Res* 2(1): 47-51. <https://doi.org/10.18689/mjpr-1000108>.
167. Zargar BA, Masoodi MH, Ahmed B, Ganie SA (2011). Phytoconstituents and therapeutic uses of *Rheum emodi* wall. ex Meissn. *Food Chemistry* 128(3): 585-589. <https://doi.org/10.1016%2Fj.foodchem.2011.03.083>.
168. Malik MA, Bhat SA, Fatima BILQUEES, Ahmad SB, Sidiqui S, Shrivastava PURNIMA (2016). *Rheum emodi* as valuable medicinal plant. *Int J Gen Med Pharm* 5(4): 35-44.
169. Sanjay Kumar, Reshma Kumari (2021) Pharmacological Activities of *Ballota nigra* (L.) Benth: A Mini Review, *International Journal of Pharma Medicine and Biological Sciences* 10(3) <http://dx.doi.org/10.18178/ijpmb.10.3.114-119>.

170. Pang Y, Wang D, Fan Z, Chen X, Yu F, Hu X, Yuan L (2014). *Blumea balsamifera*—A phytochemical and pharmacological review. *Molecules* 19(7): 9453-9477. <https://doi.org/10.3390/molecules19079453>
171. Widhiantara IG, Jawi IM (2021). Phytochemical composition and health properties of Sembung plant (*Blumea balsamifera*): A review. *Veterinary World* 14(5): 1185. <https://doi.org/10.14202/vetworld.2021.1185-1196>.
172. Al-Snafi, AE (2018) The chemical constituents and pharmacological effects of *Foeniculum vulgare*-A review. *IOSR Journal of Pharmacy* 8(5): 81-96.
173. M.J Ruffa, G Ferraro, M.L Wagner, M.L Calcagno, R.H Campos, L Cavallaro, (2002) Cytotoxic effect of Argentine medicinal plant extracts on human hepatocellular carcinoma cell line. *Journal of Ethnopharmacology* 79(3): 335-339,; [https://doi.org/10.1016/s0378-8741\(01\)00400-7](https://doi.org/10.1016/s0378-8741(01)00400-7).
174. Grassi-Zampieron R, França LV, Carollo CA, Vieira MDC, Oliveros-Bastidas A, Siqueira JMD (2010). Comparative profiles of *Achyroclinealata* (Kunth) DC. and *A. satyroides* (Lam.) DC., Asteraceae, applying HPLC-DAD-MS. *Revista Brasileira de Farmacognosia* 20: 575-579.
175. Shimizu MT, Bueno LDJF, Rodrigues RFO, Sallowicz FA, Sawaya ACHF, Marques MOM (2006). Essential oil of *Lithraeamolleoides* (Vell.): chemical composition and antimicrobial activity. *Brazilian Journal of Microbiology* 37: 556-560. <https://doi.org/10.1590/S1517-83822006000400028>
176. Silva-Júnior EF, Aquino PGV, Santos-Júnior PFS, Nascimento IJS, Gomes EA, Silva ALL, Araújo-Júnior JX (2015) Phytochemical compounds and pharmacological properties from *Schinus molle* Linnaeus and *Schinus terebinthifolius* Raddi (Anacardiaceae). *Journal of Chemical and Pharmaceutical Research* 7(12): 389-393.
177. Newman Osafo, Kwesi Boadu Mensah, Oduro Kofi Yeboah (2017) Phytochemical and Pharmacological Review of *Cryptolepis sanguinolenta* (Lindl.) Schlechter, *Advances in Pharmacological Sciences* 2017(3): 026370 <https://doi.org/10.1155/2017/3026370>.
178. Domfeh SA, Narkwa PW, Quaye O, Kusi KA, Awandare GA, Ansah C, Mutocheluh M (2021). Cryptolepine inhibits hepatocellular carcinoma growth through inhibiting interleukin-6/STAT3 signalling. *BMC complementary medicine and therapies* 21(1): 1-9. <https://doi.org/10.1186/s12906-021-03326-x>
179. Munakarmi S, Chand L, Shin HB, Hussein UK, Yun BS, Park HR, Jeong YJ (2020) Anticancer effects of *Poncirus fructus* on hepatocellular carcinoma through regulation of apoptosis, migration, and invasion. *Oncology Reports* 44(6): 2537-2546. <https://doi.org/10.3892/or.2020.7790>
180. Zhao J, Liu L, Wan Y, Zhang Y, Zhuang Q, Zhong X, Hong Z, Peng J (2015) Inhibition of hepatocellular carcinoma by total alkaloids of *Rubus alceifolius* Poir involves suppression of hedgehog signaling. *Integrative Cancer Therapies* 14(4): 394-401 <https://doi.org/10.1177/1534735415583553>
181. Ahmadiankia N (2019) Molecular targets of pomegranate (*Punica granatum*) in preventing cancer metastasis. *Iranian Journal of Basic Medical Sciences* 22(9): 977 <https://doi.org/10.22038/ijbms.2019.34653.8217>.
182. Shukla B, Saxena S, Usmani S, Kushwaha P (2021) Phytochemistry and pharmacological studies of *Plumbago zeylanica* L.: a medicinal plant review. *Clinical Phytoscience* 7: 1-11. <https://doi.org/10.1186/s40816-021-00271-7>
183. Wang X, Peng P, Pan Z, Fang Z, Lu W, Liu X (2019) Psoralen inhibits malignant proliferation and induces apoptosis through triggering endoplasmic reticulum stress in human SMMC7721 hepatoma cells. *Biological Research* 52(1): 1-13. <https://doi.org/10.1186/s40659-019-0241-8>
184. Zhao T, Sun Q, Marques M, Witcher M (2015) Anticancer properties of *Phyllanthus emblica* (Indian gooseberry). *Oxidative medicine and cellular longevity* 2015. <https://doi.org/10.1155/2015/950890>
185. Xue B, Zhao B, Luo S, Wu G, Hui X (2022) Inducing apoptosis in human hepatocellular carcinoma cell lines via Nrf2/HO-1 signalling pathway of blueberry and blackcurrant powder manipulated oat bran paste extracts. *Journal of Functional Foods* 89: 104967. <https://doi.org/10.1016/j.jff.2022.104967>.
186. Hasan MR, Alotaibi BS, Althafar ZM, Mujammi AH, Jameela J (2023) An Update on the Therapeutic Anticancer Potential of *Ocimum sanctum* L.: "Elixir of Life". *Molecules* 28(3): 1193. <https://doi.org/10.3390/molecules28031193>.
187. Kumar P, Patel D (2023) *Ocimum sanctum*: an all-round treatment for cancer?. *Alternative Therapies in Health and Medicine* 29(4): 253-257 PMID: 34331753.
188. Wang X, Peng P, Pan Z, Fang Z, Lu W, Liu X (2019) Psoralen inhibits malignant proliferation and induces apoptosis through triggering endoplasmic reticulum stress in human SMMC7721 hepatoma cells. *Biological Research* 52(1): 1-13. <https://doi.org/10.1186/s40659-019-0241-8>
189. Rajakrishnan R, Lekshmi R, Benil PB, Thomas J, Al-Farhan AH, Rakesh V, Khalaf S (2017) Phytochemical evaluation of roots of *Plumbago zeylanica* L. and assessment of its potential as a nephroprotective agent. *Saudi Journal of Biological Sciences* 24(4): 760-766. <https://doi.org/10.1016/j.sjbs.2017.01.001>.
190. Pandey P, Mehta R, Upadhyay R (2013) Physico-chemical and preliminary phytochemical screening of *Psoralea corylifolia*. *Archives of Applied Science Research* 5(2): 261-265.
191. Hasan AM, Redha AA, Mandeel Q (2018) Phytochemical investigations of pomegranate (*Punica granatum*) rind and aril extracts and their antioxidant, antidiabetic and antibacterial activity. *Nat. Prod. Chem. Res* 6(4): 332. DOI: 10.4172/2329-6836.1000332.
192. Ikram A, Khalid W, Aziz M, Arif MA, Jha RP, Khalid MZ, Izza C, Mehmood MZ, Haseeb M, Rahim MA, Naem S, Sultana F (2021) Nutritional and biochemical composition of amla (*Embllica officinalis*) and its therapeutic impact: A review. *Acta Scientific NUTRITIONAL HEALTH (ISSN: 2582-1423)* 5(2). <http://dx.doi.org/10.31080/ASNH.2020.05.0821>.
193. Chimbwali L (2022) Phytochemical Analysis of *Gossypium Hirsutum* Root Extracts Obtained From Monze District of Southern Province, Zambia-Using

- Qualitative Methods. Research Squar; <http://dx.doi.org/10.21203/rs.3.rs-1482048/v1>
194. Zhao C, Wei M, Zheng Y, Tao W, Lv Q, Wang Q, Wang S, Chen Y (2021) The Analyses of Chemical Components From *Oldenlandia hedyotidea* (DC.) Hand.-Mazz and Anticancer Effects in vitro. *Frontiers in Pharmacology* 12: 624296 <https://doi.org/10.3389/fphar.2021.624296>.
 195. Chen H, Shang X, Yuan H, Niu Q, Chen J, Luo S, Li W, Li X (2022) Total flavonoids of *Oldenlandia diffusa* (Willd.) Roxb. suppresses the growth of hepatocellular carcinoma through endoplasmic reticulum stress-mediated autophagy and apoptosis. *Frontiers in Pharmacology* 13: <https://doi.org/10.3389/fphar.2022.1019670>.
 196. Hemanth Ragav NV, Selvaraj J, Archana Santhanam DVV, Gayathri R (2021) An Update on Therapeutic Role of *Ballota Nigra* (Black Harehound)-A Review. *Annals of the Romanian Society for Cell Biology* 648-661.
 197. Samad NA, Nur N, Nik S, Kamal M, Yahaya N, Bin MY (2018) Ethnobotanical, phytochemical, and pharmacological aspects of *Melastoma* sp. *Malaysian J. Med. Heal. Sci* 14: 153-163.
 198. Yang Y, Ju Z, Yang Y, Zhang Y, Yang L, Wang Z (2021) Phytochemical analysis of *Panax* species: a review. *Journal of Ginseng Research* 45(1): 1-21. <https://doi.org/10.1016%2Fj.jgr.2019.12.009>.
 199. Huu Tung N, Uto T, Morinaga O, Kim YH, Shoyama Y (2012) Pharmacological effects of ginseng on liver functions and diseases: a mini review. *Evidence-Based Complementary and Alternative Medicine* 2012. <https://doi.org/10.1155%2F2012%2F173297>.
 200. Liu Z, Ma H, Lai Z (2021) Revealing the potential mechanism of *Astragalus membranaceus* improving prognosis of hepatocellular carcinoma by combining transcriptomics and network pharmacology. *BMC Complementary Medicine and Therapies* 21(1): 1-10. <https://doi.org/10.1186%2Fs12906-021-03425-9>.
 201. Ghasemian-Yadegari J, Hamedeyazdan S, Nazemiyeh H, Fathiazad F (2019) Evaluation of phytochemical, antioxidant and antibacterial activity on *Astragalus chrysochrys* boiss. roots. *Iranian Journal of Pharmaceutical Research: IJPR* 18(4): 1902. <https://doi.org/10.22037%2Fijpr.2019.1100855>
 202. Mishra T, Chandra P, Kumar B, Baleshwar M, Joshi P, Rana TS, Upreti DK, & Pal M (2021). Phytochemical profiling of the stem bark of *Betula utilis* from different geographical regions of India using UHPLC-ESI-MS/MS. *Analytical Science Advances* 2(11-12): 497-504. <http://dx.doi.org/10.1002/ansa.202000073>.
 203. Mishra T, Arya RK, Meena S, Joshi P, Pal, M., Meena B, Upreti DK, Rana TS, Datta D. (2016). Isolation, characterization and anticancer potential of cytotoxic triterpenes from *Betula utilis* bark. *PloS one*, 11(7): e0159430. <https://doi.org/10.1371/journal.pone.0159430>.
 204. Almeleebia TM, Alsayari A, Wahab S (2022) Pharmacological and Clinical Efficacy of *Picrorhiza kurroa* and Its Secondary Metabolites: A Comprehensive Review. *Molecules* 27(23): 8316. <https://doi.org/10.3390/molecules27238316>.
 205. Tiwari P, Mishra KP (2023) Role of Plant-Derived Flavonoids in Cancer Treatment. *Nutrition and Cancer* 75(2): 430-449. <https://doi.org/10.1080/01635581.2022.2135744>
 206. Akinyede KA, Oyewusi HA, Hughes GD, Ekpo OE, Oguntibeju OO (2022). In vitro evaluation of the anti-diabetic potential of aqueous acetone *helichrysum petiolare* extract (AAHPE) with molecular docking relevance in diabetes mellitus. *Molecules* 27(1): 155. <https://doi.org/10.3390%2Fmolecules27010155>
 207. Oyewusi HA, Wu YS, Safi SZ, Wahab RA, Hatta MHM & Batumalaie K (2022) Molecular dynamics simulations reveal the inhibitory mechanism of Withanolide A against α -glucosidase and α -amylase. *Journal of Biomolecular Structure and Dynamics* 1-16. <http://dx.doi.org/10.1080/07391102.2022.2104375>.
 208. Mustafa G, Younas S, Mahrosh HS, Albeshr MF & Bhat EA (2023) Molecular Docking and Simulation-Binding Analysis of Plant Phytochemicals with the Hepatocellular Carcinoma Targets Epidermal Growth Factor Receptor and Caspase-9. *Molecules* 28(8): 3583. <https://doi.org/10.3390/molecules28083583>.
 209. Kattan SW, Nafie MS, Elmgeed GA, Alelwani W, Badar M & Tantawy MA (2020) Molecular docking, anti-proliferative activity and induction of apoptosis in human liver cancer cells treated with androstane derivatives: Implication of PI3K/AKT/mTOR pathway. *The Journal of Steroid Biochemistry and Molecular Biology* 198: 105604. <https://doi.org/10.1016/j.jsbmb.2020.105604>.
 210. Mumtaz MZ, Kausar F, Hassan M, Javaid S & Malik A (2021) Anticancer activities of phenolic compounds from *Moringa oleifera* leaves: in vitro and in silico mechanistic study. *Beni-Suef University Journal of Basic and Applied Sciences* 10: 1-11. <https://doi.org/10.1186/s43088-021-00101-2>
 211. Siddiqui S, Upadhyay S, Ahmad I, Hussain A & Ahamed M (2021) Cytotoxicity of *Moringa oleifera* fruits on human liver cancer and molecular docking analysis of bioactive constituents against caspase-3 enzyme. *Journal of Food Biochemistry* 45(5): e13720. <https://doi.org/10.1111/jfbc.13720>.
 212. Apeh VO, Asogwa E, Chukwuma FI, Okonkwo OF, Nwora F & Uke R (2020) Chemical analysis and in silico anticancer and anti-inflammatory potentials of bioactive compounds from *Moringaoleifera* seed oil. *Advances in Traditional Medicine* 1-16. <http://dx.doi.org/10.1007/s13596-020-00521->.
 213. Hamed AN, Abouelela ME, El Zowalaty AE, Badr MM & Abdelkader MS (2022) Chemical constituents from *Carica papaya* Linn. leaves as potential cytotoxic, EGFR wt and aromatase (CYP19A) inhibitors; a study supported by molecular docking. *RSC advances* 12(15), 9154-9162. <https://doi.org/10.1039%2Fd1ra07000b>.