Inhibition of Bacterial Adhesion on Mice Enterocyte by the Hemagglutinin Pili Protein 12.8 kDa Klebsiella Pneumoniae Antibody

Dini Agustina1*, Sumarno Retoprawiro2, Noorhamdani AS2

1Department of Microbiology, Faculty of Medicine, University of Jember, Jember, Indonesia
2Department of Microbiology, Faculty of Medicine, Brawijaya University, Malang, Indonesia

ABSTRACT

Klebsiella pneumoniae as one of the most common causes of Ventilator Associated Pneumoniae is also the second most common cause of both community and hospital acquired gram negative bloodstream infections. The process of bacterial infection begins with bacterial adhesion to the host cell mediated by pili or outer membrane protein. There has not been any reported research on the hemagglutinin pili protein of K. pneumoniae as adhesion factors in VAP cases. This study was conducted in order to determine the hemagglutinin pili protein of K. pneumoniae, polyclonal antibody produced from pili protein immunization, and its ability to inhibit K. pneumoniae adhesion in mice enterocytes in VAP cases. Adhesion inhibition test used HA antibody with the implementation of dose dilutions of 1/100, 1/200, 1/400, 1/800, 1/1600, 1/3200 and 0 (control); while the immunocytochemistry test used HA pili protein with the implementation of dose dilutions of 1/10000, 1/20000, 1/40000, 1/80000, 1/160000, 1/320000 and 0 (control). Hemagglutinin pili protein was found in K. pneumoniae having MW 12.8 kDa. Pearson correlation analysis of the adhesion test showed that there was a significant correlation between antibody dilution titer with bacterial adhesion (p= 0.032, R = -0.797). Furthermore, Anova analysis of IT showed that there were significant differences between the various dilution titer with antigen-antibody reaction (p= 0.000). Antibody of hemagglutinin pili protein with MW 12.8 kDa of K. pneumoniae can inhibit the adhesion of K. pneumoniae to the enterocytes of mice in VAP cases.

Keywords: adhesion molecule, antibody, adhesion test, immunocytochemistry, K. pneumoniae

INTRODUCTION

Klebsiella pneumoniae is a member of the family of Enterobacteriaceae bacteria which is a gram negative, rod-shaped, non-motile, capsulated bacteria and facultative anaerobes [1]. K.pneumoniae can cause a variety of infections that usually attack the respiratory system and the urinary tract such as pneumonia and UTI [2]. In addition, it is also the second most common cause of community and hospital acquired gram negative bloodstream infection, 27% is noso-comial, 43% is healthcare associated community onset, and 30% is community acquired. The fatality case rate is 20%, and the annual population mortality rate is 1.3 per 100,000 and as one of the most common cases of VAP [3,4]. K.pneumoniae is widely reported to have antibiotic resistance, so the treatment for infection by these bacteria is very limited [5].

There are three pathogenicity factors of K.pneumoniae including the polysaccharide capsule, adhesion factors and lipopolysaccharide (LPS) [6]. The process of infection caused by direct contact with infectious agents begins with the host cell adhesion process either by pili or by afimbria adhesin (AFA) [7]. Gram negative bacterial adhesin played by a protein that is able to agglutinate erythrocytes of mammals known as hemagglutinin protein. Some examples of gram negative adhesion protein that has been found is the adhesin protein 36 kDa and 48 kDa on S. typhi, 16 kDa protein in A. baumannii, Outer Membrane Protein (OMP) on P. mirabilis 35 kDa, 38.19 kDa protein pili on P. aeruginosa, pili protein of 37.8 kDa on V. cholera, and OMP (Outer Membrane Protein) with MW 20kDa of K. pneumoniae [7-14]. There has not been any reported research on the hemagglutinin protein...
of pili \emph{K. pneumoniae} as an adhesion factor in VAP cases. Therefore, this study was conducted in order to determine the molecular weight of the hemagglutinin pili protein of \emph{K. pneumoniae}, polyclonal antibody produced from pili protein immunization, and its ability to inhibit \emph{K. pneumoniae} adhesion on mice enterocytes in VAP cases.

**MATERIALS AND METHODS**

This experiment used some methods, including: bacterial identification and culture, isolation of bacterial pili, isolation of bacterial hemagglutinin pili protein, hemagglutination test, and production of polyclonal antibody, serum collection of mice, adhesion inhibition test, and immunocytochemistry test. These methods were used to determine which hemagglutinin pili protein that would be used in adhesion inhibition test and immunocytochemistry test.

**Culture of \emph{Klebsiella pneumoniae}**

Samples were taken from bronchial aspirate specimens of VAP patients admitted to the Intensive Care Unit during the period of January to June 2013 from Clinical Microbiology laboratory of Saiful Anwar Public Hospital, Malang. Identification of the bacteria was performed according to the instructions by Ling et al. and Sikarwar & Batra. Bacteria were grown in TCG medium. The growing culture was collected by scraped, previously poured sufficient sterile PBS pH 7.4. The bacterial suspension was put in a bottle containing of 1000 ml of Brain Heart Infusion Broth (BHI). The suspension was shaken for 30 minutes on a powerful water bath at 37°C. Bacterial suspension was taken as much as 10 ml and put in TCG medium and incubated for 48 hours at 37°C [15-17].

**Isolation of \emph{K.pneumoniae} hemagglutinin pili protein (SDS-PAGE)**

Determination of protein’s molecular weight was done by using SDS-PAGE. Protein samples were heated at 100°C for 5 min in a buffer solution containing 5 mM Tris HCl pH 6.8, 2-mercaptoethanol 5%, Sodium Dodecyl Sulfate 2.5% w/v, glycerol 10% v/v with color tracer Bromophenol blue. Separating gel concentration chosen was 12.5% mini slab gel with a 3% stacking gel. 120 mV and 400 mA voltage was used, with a running time of over 90 minutes. Comassic Brilliant Blue R-250 was used as a dye and as a pre-stained broad range protein marker [18].

**Hemagglutination test**

Hemagglutination test was performed to determine which pili cut had the highest hemagglutination titer to be used in the next protocol. Hemagglutination test was done according to the instructions of Li. Serial dilutions of samples were made for each dilution and the volume of each micro plate well was 50 µl. A suspension of red blood cells of mice with a concentration of 0.5% was added to each sample with the same volume as much as 50 µl. Then it was shaken with rotator plate for 1 minute, and then subsequently placed in room temperature for 1 hour. The magnitude of the titer was determined by observation on the presence of agglutination of red blood cells on the lowest dilution [19].

**Production of polyclonal antibodies**

Mice used in this study was strain BALB/C female mice, aged 6-8 weeks. Antigens used were previously selected hemagglutinin pili protein of \emph{K. pneumoniae}. The antigen in the syringe was emulsified with Complete Freund's Adjuvant (CFA). Mice were injected intraperitoneally with a dose of 50 µg antigen diluted in normal saline solution. Booster injection was performed in week two, three and four by using antigen emulsified with Incomplete Freund's Adjuvant (IFA) with the same dose. Serum will be taken ten days after the last booster [20-22].

**Serum collection method**

Blood was collected from the heart. Blood was taken from five mice, after that it was collected in sterile tubes and placed into the
incubator with temperature of 37°C in a tilted position for 30 minutes. Then it was stored in a refrigerator with the temperature of 4°C for 10 min and then centrifuged at 10,000 rpm for 5 minutes. Supernatants were taken and put in sterile tubes and stored at -20°C [23].

Isolation of mice enterocyte cells
Mice enterocyte were used as a model of bacterial adhesion on host cell. Isolation of enterocytes cells of mice was performed according to the Weisser method. Mice used were healthy mice with approximate weight of 25 grams. Mice were anesthetized using chloroform and then parts of the small intestine were cut and taken. The small intestine was washed with PBS pH 7.4 containing 1 mM dithiothretiol at 4°C until it looked clean. Parts of small intestine then put in the fluid containing of 1.5 mM KCl, 9.6 mM NaCl, 27 mM NA citrate, 8 mM KH2PO4 and 5.6 mM Na2HPO4 with pH 7.4, then incubated in shaking incubator for 15 minutes, with a temperature of 37ºC. Supernatant was discarded and the tissue was transferred to a liquid containing 1.5 mM EDTA and 0.5 mM dithiothretiol. It was then shaken strongly for 15 minutes. Supernatants were taken and put in sterile tubes and stored at -20ºC [23].

Adhesion inhibition test
The adhesion test used a modified method of Nagayama et al. K. pneumoniae cultured in lactose broth at 37ºC. Furthermore, the bacteria were harvested using centrifugation of 6,000 rpm for 10 min at 4ºC. The precipitate was suspended with PBS and bacteria content were made 10⁶ cells/ml using a spectrophotometer at a wavelength of 560 nm to a concentration of 10⁴ cells/ml. Enterocytes were ready to be used in adhesion test [24].

Gram staining
Staining was performed to gain main description of the morphology of enterocytes and bacterial adhesion of K. pneumoniae on enterocytes. Slide was protected using the crystal violet for 1 min and rinsed with water. Later, lugol was dropped for 1 minute followed with 96% ethyl alcohol washing. Furthermore, safranin was dropped for 30 seconds and the slide was rinsed with water. The observation was done under the microscope with a magnification of 1000x.

Immunocytochemistry test
Prior to the immunocytochemistry procedure, sample preparations of hemagglutinin pili protein on mice enterocytes were performed as previously described in the methods. Samples were then fixed with methanol and washed 3 times with PBS pH 7.4 for 5 minutes, then given a 3% H2O2 for 15 min and washed again with PBS pH 7.4 3x5 minutes. Blocking was done with triton X-100 (0.25%) in BSA blocking buffer for 1 h incubation at room temperature, washed with PBS pH 7.4 3x5 minutes, subsequently incubated for 24 h at 4ºC with primary antibodies (polyclonal antibody hemagglutinin pili protein) 1:100 with BSA blocking buffer, washed with PBS pH 7.4 3x5 minutes. The next step was incubation with secondary antibody anti-mouse IgG (1:200) for 1 h, washed again with PBS pH 7.4 3x5 minutes. Then the SA-HRP (1:500) was dropped in each preparation, incubated for 40 min, washed with PBS pH 7.4 for 3x5 minutes and distilled water for 2x5 minutes. After that, it was incubated with di-aminobenzine (DAB) for 20 min, washed again with PBS pH 7.4 for 3x5 minutes and distilled water for 2x5 minutes. Mayer hematoxylin was last given for 10 minutes, covered with a cover glass and spilled entelan. The results was ready to be observed under 400-1000x [9,25] microscope magnification.

RESULTS AND DISCUSSION
Hemagglutinin protein is a protein that is able to agglutinate mammalian erythrocytes. The existence of the hemagglutinin protein in bacteria is a marker that the bacteria have the ability to perform the process of adhesion, an important process in the initiation and progression of clinical symptoms of a disease. The sample used in this study had the highest hemagglutination titer (data not shown) [7,26].

The result of pili protein identification by SDS-PAGE method can be seen in Figure 1. Four pili cutting were obtained and analyzed with SDS-PAGE; the profile of subunit pili protein from Pili I, Pili II, Pili III, and Pili IV were identical. Some bands that appeared in fourth pili cutting were also appeared in whole...
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cells but not in the bare cell. It means that those seven bands were presented only in pili (Figure 1).

Figure 1. Profile of subunit pili protein of \textit{K. pneumoniae} (SDS-PAGE 12.5%). 1: Marker, 2: Pili I, 3: Pili II, 4: Pili III, 5: Pili IV, 6: Bare cell (20x dilution), 7: Bare cell (50x dilution), 8: Whole cell (20x dilution). There were seven bands presented only in pili (91.2 kDa, 42.4 kDa, 36 kDa, 27.4 kDa, 20.9 kDa, 12.8 kDa, and 10.8 kDa)

Four pili cutting were then performed the hemagglutination test to determine which pili cutting to be used in the next step. The result showed that Pili I had the highest hemagglutination titer (1/256 titer) (data not shown). Seven bands in Pili I were then performed the hemagglutination test. The results can be seen in Figure 2.

Figure 2. Profile of hemagglutination test of subunit pili protein of \textit{K. pneumoniae}. 1: 91.2 kDa, 2: 42.4 kDa, 3: 36kDa, 4: 27.4 kDa, 5: 20.9kDa, 6: 12.8 kDa, 7: 10.8 kDa, and K: control. Subunit pili protein with MW 12.8 kDa (row no.6) showed the highest HA titer (1/256 titer).

The result were then confirmed by spectrophotometer and showed the same result, in which subunit pili protein with MW 12.8 kDa had the highest concentration at 1.0779 mg/ml (data not shown). Thus, the most likely factor in bacterial adhesion was played by pili protein of 12.8 kDa.

Based on the result, subunit pili protein 12.8 kDa were selected to produce polyclonal antibody in mice. The antibody obtained from immunization of mice was then used in adhesion inhibition test. This test used various concentration of antibody which were 1/100, 1/200, 1/400, 1/800, 1/1600, 1/3200 and 0 (control) (p<0.05) (Figure 3).

Figure 3. Results of adhesion inhibition test of \textit{K. pneumoniae} into enterocytes after coated with various concentration of polyclonal antibody. Increased dose dilutions were resulted in increased bacterial adhesion into enterocytes cells.

Based on the result showed in Figure 3, the amount of bacteria attached into enterocytes could be calculated. The adhesion index (AI) was calculated by counting the number of bacteria attached to 100 enterocytes (Table 1). Pearson correlation analysis was performed based on the observation of AI. The result is shown in Table 1.

Table 1. The correlation between various concentration of polyclonal antibody with Adhesion Index.

<table>
<thead>
<tr>
<th>group</th>
<th>N</th>
<th>Mean ± SD</th>
<th>p-value</th>
<th>Pearson correlation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Titer 1/100</td>
<td>3</td>
<td>116 ± 14.42</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Titer 1/200</td>
<td>3</td>
<td>113.33 ± 20.01</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Titer 1/400</td>
<td>3</td>
<td>135.67 ± 10.02</td>
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</tr>
<tr>
<td>Titer 1/1600</td>
<td>3</td>
<td>280 ± 18.68</td>
<td>0.032</td>
<td>-0.797</td>
</tr>
<tr>
<td>Titer 1/3200</td>
<td>3</td>
<td>289 ± 30.55</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>3</td>
<td>293.33 ± 27.43</td>
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</table>
The ability of polyclonal antibodies (immuno-globulin G) against pili protein MW 12.8 kDa to inhibit bacterial adhesion was then tested by the adhesion inhibition test. The results indicate that the adhesion of K. pneumoniae bacteria was inhibited by polyclonal immuno-globulin G resulting from the pili protein immunization (Figure 3). As seen in Figure 3, the lowest dilution (1/10,000 titer) resulted in the smallest amount of bacteria that attached to the enterocytes cells of mice, then in the next dilution the number of bacteria attached were increasing. This is caused by the ability of the circulating antibodies to pili bacteria to bind actively to bacterial pili so the bacteria cannot attach to the enterocytes [27,28]. Based on the result of adhesion inhibition test using an antibody of subunit pili protein K. pneumoniae MW 12.8 kDa, it can be concluded that the polyclonal antibody is an antibody that can fight against an adhesion molecule. Similar results are obtained in a study conducted by Agustina et al. proving that the polyclonal antibodies obtained from the injection of 49.8 kDa subunit pili of S. dysentriae in mice acts as an antibody against adhesion molecules [29].

Based on the result of Pearson correlation test, there was a significant correlation between antibody dilution titer with adhesion index (p=0.032) with a correlation coefficient of -0.797 (Table.1). This means that the dose of antibodies affect adhesion index as much as 79.7 %. Decreased antibodies concentration would results in increased bacterial adhesion index. These results are similar to the results of the study conducted by Hidayati revealing that in the adhesion of Pseudomonas aeruginosa pili, decreased antibodies concentration will increase the bacterial adhesion index significantly [31]. Similar results have also been obtained by other studies [9,27,30].

Of the three bacterial attachment patterns (local, aggregative, and diffuse), the pattern of bacterial attachment of K. pneumoniae in this study could be concluded to be more likely to the diffuse type. This can be seen by looking at the results of the adhesion test in Figure 3. It appears that the attachment of K. pneumoniae bacteria was evenly distributed on the enterocytes cells. This attachment is mediated by the hemagglutinin pili protein MW 12.8 kDa of K. pneumoniae. Previous research conducted by Di Martino et al. proves the role of nonfimbrial protein K. pneumoniae with MW 29 kDa as an intermediary diffuse type of bacterial adhesion to Caco-2 cells [31].

The second method used to ensure that the subunit pili protein MW 12.8 kDa of K.pneumoniae is an adhesion molecule was immunocytochemistry test. Adhesion molecules can attach to receptor molecules and the receptors are part of the cell [29]. In an immunocytochemistry test, antigen and antibody reactions can be evaluated from the color that appears. Brown color indicates that there is conformity antigen-antibody binding, while the blue color indicates a discrepancy between the two [9].

The concentration of antibody used in this test was 1/100, based on the strongest antigen-antibody reaction by Dot Blot method (data not shown). The effect of various concentration of subunit pili protein with MW 12.8 kDa of K.pneumoniae in immunocytochemistry test (IT) will produce color in enterocyte. The result showed the brown and blue color (Figure 4).

Brown color indicates a positive result, it means that there is a perfect bond of antigen and antibody, while the blue color indicates that the antigen-antibody bond is not perfect. Brown enterocytes cells were calculated to 100 entero-cytes and repeated for three times. Statistical analyses employing a One Way ANOVA and Tukey post hoc analysis were done in order to determine whether there are differences between the various dilution titers with antigen-antibody reaction. The results can be seen in Table 2. Results of One Way Anova analysis showed that there were significant differences between the various dilution titer with antigen-antibody reaction (p=0.000) (95% confidence level). The results of comparative analysis of each group through post hoc test (Tukey) can be seen in Table 2.

In Figure 4, it appeared that the lowest dilution of antigen (1/10000) showed a positive result (brown color) in almost all enterocytes cells, then by increasing dilution (decreased concentration) brown color that appeared decreased until blue colors fully appeared in control. Many antibodies
of hemagglutinin pili protein bound to the receptors appeared at the lowest dilution, so the brown color produced were also many. Appearance of color gradation changes from brown to blue along with the decreased of pili proteins that bind to these antibodies can be seen in Figure 4 (1/20000-1/320000 titer). These results demonstrate the hemagglutinin pili protein MW 12.8 kDa of *K.pneumoniae* really an adhesion protein that has the ability to bind enterocytes cells. These results are similar to previous studies that the pili pro-tein is an adhesion molecule via immuno-cytochemistry test [9,29]. Based on Table 2, there was a significant difference between the antigen-antibody reaction with various dilution titers of hemagglutinin pili protein MW 12.8 kDa of *K.pneumoniae* (p= 0.000 with a confidence interval (α)= 95%).

**Table 2. The relationship between various dilution titers with antigen-antibody reaction using immunocytochemistry test**

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Mean ± SD</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Titer 1/10000</td>
<td>3</td>
<td>93.00 ± 3.82</td>
<td></td>
</tr>
<tr>
<td>Titer 1/20000</td>
<td>3</td>
<td>85.00 ± 5.57</td>
<td></td>
</tr>
<tr>
<td>Titer 1/40000</td>
<td>3</td>
<td>80.33 ± 4.16</td>
<td></td>
</tr>
<tr>
<td>Titer 1/80000</td>
<td>3</td>
<td>68.33 ± 3.06</td>
<td>0.000</td>
</tr>
<tr>
<td>Titer 1/160000</td>
<td>3</td>
<td>61.33 ± 8.50</td>
<td></td>
</tr>
<tr>
<td>Titer 1/320000</td>
<td>3</td>
<td>38.67 ± 2.52</td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>3</td>
<td>0.00 ± 0.00</td>
<td></td>
</tr>
</tbody>
</table>

Description: The different notation means significant difference (p<0.05) and when it contains the same notation it means no significant difference (p> 0.05).

This study was the first which characterized pili protein of *K. pneumoniae* taken from bronchial aspirates of VAP cases. This identified pili protein can be used to develop a diagnostic kit and a vaccine against *K. pneumoniae*. It could help clinicians to treat their patients admitted to the Intensive Care Unit so it can reduce morbidity and mortality because of Ventilator Associated Pneumoniae caused by *K. pneumoniae*. Different specimen from the same bacteria could result in different pili protein to be identified, as proven by the research conducted by Sukarjati using hemagglutinin pili protein MW 32.23 kDa of *E.coli* taken from semen specimen of infertile man, and another study by Sudana et al. using hemagglutinin pili protein MW 37 kDa of *E. coli* taken from midstream urine of UTI patients [32,33]. Further researches are needed to examine hemagglutinin pili protein of *K.pneumoniae* taken from other specimen and other cases.

**CONCLUSION**

Based on the results and discussion, it can be concluded that antibodies of hemagglutinin pili protein with MW 12.8 kDa of *K. pneumoniae* can act as an adhesion molecule. Through adhesion inhibition and immunocytochemistry test, it has been found out that hemagglutinin pili protein with MW 12.8 kDa of *K. pneumoniae* antibody can inhibit the adhesion of *K. pneumoniae* to the enterocytes of mice. The need to do further research is to find the characters of molecular sub-units in pili adhesion protein of *K. pneumoniae* found in other isolates and other cases apart from VAP.

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**REFERENCES**


